

Fatty Alcohols (Octanol and Decanol)

Crops

1 Identification of Petitioned Substance: Fatty Alcohols

2

3 **Chemical Names:** Octanol: InChI=1S/C8H18O/c1-2-3-4-5-6-7-8-
9/h9H,2-8H2,1H3; InChI
4 Fatty Alcohols (54.5% Decanol, 45.1% Octanol, Key=KBPLFHHGFOOTCA-UHFFFAOYSA-N;
5 0.4% Hexanol); octan-1-ol; decan-1-ol Canonical SMILES=CCCCCCCCO; EC Number=
6 **Other Name:** Octyl-Decyl Alcohol, 1-Octanol, 1 203-917-6, 271-642-9, 606-925-1; ICSC
7 Octanol, Alcohol, n-Octyl, n Octanol, n Octyl Number=1030, 1170; RTECS
8 Alcohol, n-Octanol, n-Octyl Alcohol, 1-decanol; n- Number=RH6550000, RH795000;
9 decanol; n-decyl alcohol; n-decyl alcohol, aluminum UNII=NV1779205D;
10 salt; n-decyl alcohol, magnesium salt; n-decyl alcohol, Decanol: InChI= 1S/C10H22O/c1-2-3-4-5-6-7-8-
11 sodium salt; n-decyl alcohol, titanium salt 9-10-11/h11H,2-10H2,1H3; InChI Key=
12 **Trade Names:** MWKFXSUHUHTGQN-UHFFFAOYSA-N;
Kleen-tac 85, O-Tac, Green Tac, Ten-Tac, Sucker- Canonical SMILES= CCCCCCCCCO; EC
Plucker, Alfol 80, and others. Number= 203-956-9, 266-367-6, 253-173-1, 287-
CAS Numbers: 621-2, 613-644-8; ICSC Number=1490; RTECS
68603-15-6; 112-30-1; 66455-17-2; 85566-12-7 Number=HE4375000; UN Number=3082, 1987;
UNII=89V4LX79
Other Codes:

13

14 Summary of Petitioned Use

15 A petition was received by the National Organic Program to add Fatty alcohols, a mixture of aliphatic alcohols,
16 hexanol, octanol and decanol to the National list 7 CFR 205.601 (k) under the heading plant growth regulator. The
17 petitioned use for a mixture of fatty alcohols, consisting mainly of octanol and decanol, is topping and sucker
18 control on organic crops.

19 Characterization of Petitioned Substance

20 **Composition of the Substance:**

21 Monohydric aliphatic alcohols containing six to ten carbons are called lower alcohols. Those containing twelve to
22 twenty carbons are called higher alcohols (Atwood, 1963). Historically, lower and higher alcohols were derived
23 from natural fats, oils and waxes hence the name fatty alcohols. Now a similar mixture of fatty alcohols that is
24 synthetically produced from petrochemical feedstocks, is widely available (NPCS Board of Consultants &
25 Engineers, 2010). The C₆-C₁₂ linear alcohols are used for plasticizers, surfactants, tobacco sucker control, mining
26 chemicals and in the manufacture of fatty amines.

27 ALFOL 810 Alcohol, a product containing C₆-C₁₂ linear alcohols was the first mixed fatty alcohol product
28 registered with the Environmental Protection Agency as a tobacco topping and suckering agent. It was registered
29 by the Continental Oil Company under EPA reg. no. 39496-1. ALFOL 810 has the same composition as Mascol-80
30 which is referenced in the petition with EPA registration number 63896-1 (Table 1).

31 The Alfol process to produce fatty alcohols combines aluminum, ethylene, hydrogen and air to synthetically
32 produce a mixture of lower and higher alcohols (Atwood, 1963). However, sufficient quantities of plant derived
33 raw materials (e.g. palm kernel oil) for this product are available. The Davy process is used to produce fatty
34 alcohol from vegetable oils. Fair products, Inc., mentioned in this petition, and the Drexel Chemical Company
35 currently market fatty alcohols for tobacco sucker control. Drexel's tobacco sucker control agent is called "Sucker-
36 Plucker." It contains 85% active ingredients (fatty alcohols) and 15% non-active ingredients. The non-active label
37 ingredient polysorbate-80 (tween-20) is a surfactant essential to the effectiveness of the fatty alcohols (Tso et al.,

1975). The EPA registration number for this product is 19713-35 (Table 1). It is derived from a fatty alcohol mixture typically containing C₆ 0.5%, C₈ 42.0%, C₁₀ 56% and C₁₂ 1.5% linear alcohols (EPA, 1993, 2004, 2011). A similar product to “Sucker-Plucker” from Fair products, Inc. is cited in this petition as O-TAC Plant contact agent (EPA reg. no. 51873-18) containing 36.2% Octanol, 48.2% decanol and 0.3% dodecanol (Table 1). O-TAC is currently described as a plant control agent for organic, purity residue clean (PRC) and maleic hydrazide free tobacco. Table 1 provides information about several fatty alcohol products for tobacco topping and suckering.

Fatty Alcohol Product Name	Composition on EPA registered Label (percent)				EPA Registration Number
	C ₆	C ₈	C ₁₀	C ₁₂	
ALFOL 810	≤ 1	39-47	51-59	≤ 1	39496-1
Mascol-80	-	45.1	54.5	-	63896-1
Sucker-Plucker	0.5	42.0	48.2	0.3	19713-35
O-TAC	-	36.2	48.2	-	51873-18
N-TAC		45	55		51873-18

44

45 **Source or Origin of the Substance:**

46 Petroleum derived feedstocks are the primary source material for synthetically produced medium chain
 47 length lower and higher alcohols. However, palm, palm kernel and coconut oils are also available sources
 48 for producing fatty alcohols in the C₈-C₁₄ chain lengths (Reck, 1985). Because of the importance of fatty
 49 alcohols in many industries, there is significant ongoing research in producing medium chain length fatty
 50 alcohols via fermentation in yeast or bacteria, i.e. enzymatic reduction of fatty acids. Although reports have
 51 shown positive findings, this production route is not yet available commercially. Therefore, naturally
 52 produced fatty alcohols are not yet available in sufficient quantities for commercial use as topping and
 53 suckering agents (Wang, 2016).

54 **Properties of the Substance:**

55 Fatty alcohols produced from palm oil, palm kernel oil and coconut oil by the Davy process for use as tobacco
 56 suckering agents are aliphatic alcohols with six to twelve carbons; however the most abundantly produced fatty
 57 alcohols in this mixture are caproic alcohol, caprylic alcohol, and capric alcohol (Tables 2, 3, Fig. 1). The primary
 58 alcohols exhibit some trends. For example, for each additional -CH₂- unit the normal boiling point increases
 59 about 20°C, the specific gravity increases 0.003 units and the melting point increases about 10°C in the lower
 60 range and about 4°C in the higher range. Water solubility decreases and oil solubility increases with increasing
 61 molecular weight. The fatty alcohols are all liquid with light natural fruity odors (NPSC, 2010). Both the density
 62 and viscosity of fatty alcohols increase with increasing molecular weight (Rauf et al., 1983).

63 **Specific Uses of the Substance:**

64 Fatty alcohols, octanol and decanol are used to chemically remove flower buds and suckers from tobacco
 65 plants. Removal of the flower tops and the suckers encourages the growth of larger leaves. The use of fatty
 66 alcohols is an alternative to two laborious hand operations in tobacco production. Topping or removal of
 67 buds or flowers and subsequent removal of suckers (lower leaves) by hand requires ten or more hours per
 68 acre. A course spray of 5% decanol or a combination of decanol and octanol applied before bud formation
 69 inhibits the formation of the bud. Fatty alcohol dripping down the stem of the plant inhibits sucker
 70 formation. Yields are also increased with the use of this treatment (USDA-ARS, 1970).

71

Table 2 Physical properties of the low molecular weight fatty alcohols*

IUPAC name	Common name	CAS registry number	Molecular formula	MW	Hydroxyl number	Melting point, °C	Boiling point, °C (p.kPa)
1-Hexanol	Caproic alcohol	111-27-3	C ₆ H ₁₄ O	102.2	548	-52	157
1-Heptanol	Enanthic alcohol	111-70-6	C ₇ H ₁₆ O	116.2	482	-30	176
1-Octanol	Caprylic alcohol	111-87-5	C ₈ H ₁₈ O	130.2	430	-16	195
1-Nonanol	Pelargonic alcohol	143-08-8	C ₉ H ₂₀ O	144.3	388	-4	213
1-Decanol	Capric alcohol	112-30-1	C ₁₀ H ₂₂ O	158.3	354	7	230
1-Undecanol	-	112-42-5	C ₁₁ H ₂₄ O	172.3	326	16	245
1-Dodecanol	Lauryl alcohol	112-53-8	C ₁₂ H ₂₆ O	186.3	300	23	260

from (Condea, 2000)

72 **Approved Legal Uses of the Substance:**

73 Fatty (aliphatic) alcohols containing a mixture of six, eight, ten and twelve carbons are approved by the US
 74 Environmental Protection Agency as the active ingredients for use as plant regulators for tobacco sucker
 75 control. Synthetic hexyl-, octyl- and decyl-alcohol maybe safely used in food and in the synthesis of food
 76 components provided that the alcohol's purity of the alcohol is 99% or greater. Synthetic fatty alcohols may
 77 only be used as substitutes for and to the extent that naturally derived fatty alcohols are found in foods (US
 78 FDA, 21 CFR §172.864). The US Department of Agriculture does not regulate the use of fatty alcohols for
 79 tobacco sucker control.

80 **Action of the Substance:**

81 Tobacco grades are based on the manufacturer's use for cured leaf. Grades are determined by certain
 82 physical and chemical properties. These properties are to a great extent determined by the position of the
 83 leaf on the plant. Usually leaves on the lowest part of the plant are not considered useful and removed in
 84 the suckering process (Calvert, 1956a). Tso (1964) first reported that alkyl esters of C₉ to C₁₂ fatty acids
 85 inhibited the growth of axillary buds when applied to tobacco plants that had been topped (Tso et al.,
 86 1965). It was later reported that both lower alkyl esters of the C₈ to C₁₂ fatty acids and the C₈ to C₁₀ fatty
 87 alcohols in aqueous emulsions selectively killed the terminal meristems of a wide variety of plants without
 88 damaging the axillary meristems, foliage, or stem tissues of the plants (Cathey et al., 1966). Fatty alcohols
 89 depress the surface tension along chloroplast and mitochondrial membranes reducing phosphorylation by
 90 allowing water to leak into a hydrophobic lipid-protein environment inhibiting nicotinamide adenine
 91 dinucleotide dependent respiration (Thore and Baltscheffsky, 1965). The leakiness causes desiccation
 92 producing a burned appearance on the leaf tissue surface (Wheeler et al., 1991). Fatty alcohols with chain
 93 lengths of C₉, C₁₀, and C₁₁ are better on a molar basis in selectively killing or inhibiting axillary and terminal
 94 bud growth of tobacco than the corresponding alkyl esters. Pure 1-decanol is reportedly the most effective
 95 topping and suckering agent (Steffens and McKee, 1969).

96 Without an appropriate surfactant both the C₉ to C₁₁ methyl esters and the C₈ to C₁₀ fatty alcohols exhibited
 97 nonselective tissue kill. At a concentration as low as 1.4x10⁻¹ Molar, fatty alcohols cause immediate swelling
 98 of plant cell nuclei and a general cessation of cell division. Cells can become poly-nuclear as a result of
 99 exposure. At high concentrations fatty alcohols cause discolorization of root tips (Tso and Burk, 1969).
 100 However, with a surfactant the fatty alcohol emulsions are phytotoxic only to young meristematic tissue
 101 but caused little or no visible injury to more mature tissue (Steffens et al., 1967).

102

103 Topping and desuckering influence the yield and the quality of tobacco topping at the elongated bud stage
104 with 16 leaves producing good yield and quality (Rao et al., 2003). The use of a 4% solution of 1-decanol
105 applied by brushing 10 days after topping to the ten top axils of tobacco improved yield 67% over hand
106 topping or treatments with Neem oil (Singh et al., 1998) The value of topped and suckered tobacco is
107 higher. The use of 7.5% fatty alcohols with a surfactant improves both the price and the yield of burley
108 tobacco. The alkaloid content in topped tobacco is higher than untopped, although total nitrogen was
109 unchanged (Mylonas and Pangos, 1977).

110 If manual topping is performed on tobacco immediately after flower bud formation, manual suckering
111 must be done frequently and regularly for the following three months as the axillary growth continues.
112 Applying fatty alcohols as a topping agent and/or after topping can reduce the intensive labor required for
113 topping and/or suckering (Bhat et al., 1994). Production lots of fatty alcohols contain a distribution of C₆-
114 C₁₂ chain length alcohols. The C₆ alcohols are not active suckercides. Whereas, C₈ chain length alcohol is
115 about 50% effective. C₉, C₁₀ and C₁₁ chain length alcohols are 100% effective and C₁₂ length alcohol is less
116 effective. Higher chain lengths are not effective (Steffens et al., 1967). Suckering with fatty alcohols (C₆-C₁₂)
117 generally produced a higher yield than manual suckering and does not alter USDA grading characteristics
118 (Bruns, 1987).

119 **Combinations of the Substance:**

120 Fatty alcohols are combined with the surfactant polyoxyethylene (20) sorbitan monooleate, i.e. polysorbate
121 80, (tween 20). The surfactant permits an aqueous emulsion to be formed in the fatty alcohol solution,
122 providing an even distribution of the active substance. There is clear evidence that the use of fatty alcohols
123 for tobacco topping and suckering without a surfactant causes damage to the plant (Tso, 1964). Polysorbate
124 80 is a mixture of polyoxyethylene ethers of mixed partial oleic esters of sorbitol anhydrides and related
125 compounds. It is approved by the FDA for safe use as an additive in many food applications (21 CFR
126 §172.840).

127 Status

128 **Historic Use:**

129 Topping and suckering have been used in tobacco production for many years. Benefits include improved
130 tobacco quality and a significant increase in tobacco yield (Hunter, 1954). Hand topping tobacco plants in
131 late summer promotes full development of upper leaves adjacent to the terminal (flower) buds. Two weeks
132 after topping, suckers that have subsequently developed along the stalk are also removed by hand. Suckers
133 interfere with harvesting and curing operations. As much as ten or more hours per acre are spent on hand
134 topping and suckering (USDA-ARS, 1970). Fatty alcohols were introduced in conventional tobacco
135 production as chemical topping and suckering agents in the late 1960s. The use of fatty alcohols for topping
136 and suckering also significantly increases yield (USDA-ARS, 1970).

137 **Organic Foods Production Act, USDA Final Rule:**

138 There is no reference in the National List for fatty alcohols. The proposal to add fatty alcohols to the
139 National List specifies 7 CFR 205.601 (k) under the heading plant growth regulator. This section of the
140 National List currently describes the use of the synthetic substance ethylene in organic crop production as a
141 plant growth regulator for regulation of pineapple flowering. Fatty alcohols as aqueous emulsions inhibit
142 terminal or axillary bud growth of tobacco plants. Contact with meristematic tissue affects plant
143 development by preventing the growth flower buds and suckers (Tso et al., 1965; Steffens and Barer, 1984).

144 **International**

145 **Canada - Canadian General Standards Board Permitted Substances List (CAN/CGSB-32.311-2015)**

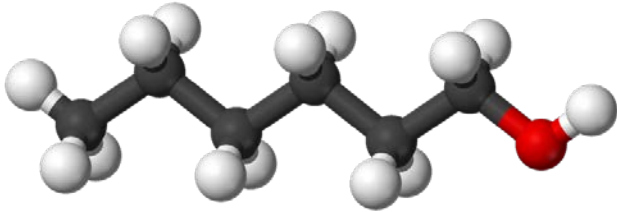
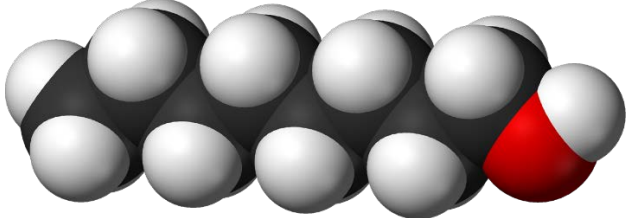
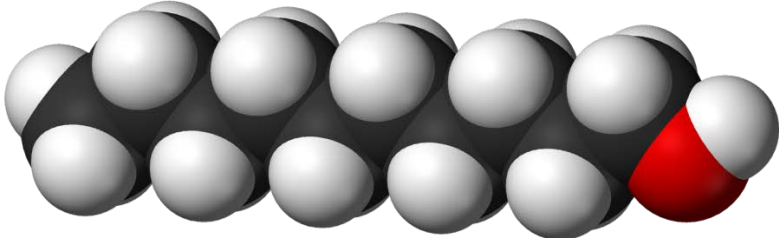
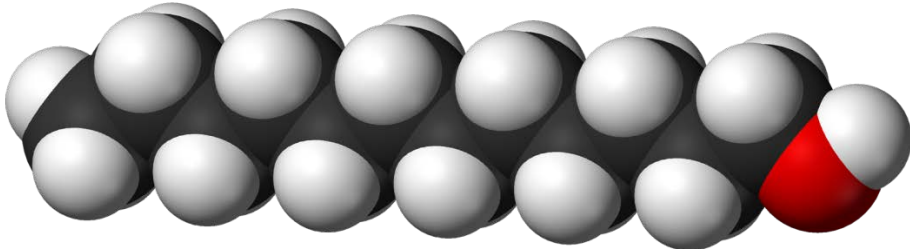
146 Fatty alcohols were not found in the Canadian standard (CAN/CGSB 2015).

147 **CODEX Alimentarius Commission, Guidelines for the Production, Processing, Labelling and Marketing** 148 **of Organically Produced Foods (GL 32-1999) -**

149 Fatty alcohols were not found in the Codex Organic guidelines (GL 32-1999).

150 **European Economic Community (EEC) Council Regulation, EC No. 834/2007 and 889/2008**

151 Fatty alcohols were not found in the EEC organic standards, EC No. 834/2007 and 889/2008.

Table 3 Space Filling Structure Renditions of Lower Fatty Alcohols*	
1-Hexanol	
1-Octanol	
1-Decanol	
1-Dodecanol	
*carbon=black, hydrogen=white, oxygen=red	

152 **Japan Agricultural Standard (JAS) for Organic Production –**

153 Fatty alcohols were not found in the MAFF organic standard.

154 **International Federation of Organic Agriculture Movements (IFOAM) –**

155 Fatty alcohols were not found in the IFOAM guidelines.

Evaluation Questions for Substances to be used in Organic Crop or Livestock Production

157 **Evaluation Question #1: Indicate which category in OFPA that the substance falls under: (A) Does the**
 158 **substance contain an active ingredient in any of the following categories: copper and sulfur**
 159 **compounds, toxins derived from bacteria; pheromones, soaps, horticultural oils, fish emulsions, treated**
 160 **seed, vitamins and minerals; livestock parasiticides and medicines and production aids including**
 161 **netting, tree wraps and seals, insect traps, sticky barriers, row covers, and equipment cleansers? (B) Is**
 162 **the substance a synthetic inert ingredient that is not classified by the EPA as inerts of toxicological**

163 **concern (i.e., EPA List 4 inerts) (7 U.S.C. § 6517(c)(1)(B)(ii))? Is the synthetic substance an inert**
164 **ingredient which is not on EPA List 4, but is exempt from a requirement of a tolerance, per 40 CFR part**
165 **180?**

166 Fatty alcohols do not fall into any of the OFPA categories. Fatty alcohols produced as a mixture of four
167 aliphatic alcohols are not considered inert by the Environmental Protection Agency nor are they included
168 in List 4. Fatty alcohols may be registered with the EPA only for tobacco sucker control. N-decyl alcohol
169 (decanol) and n-octyl alcohol (octanol) are individually approved by the US Food and Drug
170 Administration (FDA) for food and non-food use as solvents or co-solvents (7 CFR §172.864 Synthetic fatty
171 alcohols).

172 **Evaluation Question #2: Describe the most prevalent processes used to manufacture or formulate the**
173 **petitioned substance. Further, describe any chemical change that may occur during manufacture or**
174 **formulation of the petitioned substance when this substance is extracted from naturally occurring plant,**
175 **animal, or mineral sources (7 U.S.C. § 6502 (21)).**

176 Fats and oils from animal and vegetable sources (oleochemical) are the primary feedstocks for
177 manufacturing naturally sourced fatty alcohols. These feedstocks contain triglycerides composed of three
178 fatty acids and glycerol. Alcohols are produced by esterification and/or reducing the fatty acid functional
179 groups. Any triglyceride or fatty acid may be used as a raw material for fatty alcohol production (NCPS,
180 2010). The common sources are coconut oil, palm kernel oil, lard, tallow, rapeseed oil, soybean oil and corn
181 oil. Producers of natural fatty alcohols typically make a broad range of fatty alcohol products having
182 various carbon chain lengths. They vary feedstocks to meet market needs for particular alcohols and to
183 take advantage of changes in the relative costs of the various feedstock material (NCPS, 2010; Reck, 1985).

184 There are a number of production processes for producing natural fatty alcohols. These have evolved since
185 the 1930's to improve safety and efficiency. The earliest production began with the sodium reduction of the
186 methyl esters from natural fats and oils. In this process molten sodium was mixed with dried ester and
187 alcohol which acts as a hydrogen donor. Many safety precautions were necessary due to the large amount
188 of metallic sodium used. A safer hydrogenolysis process was developed to replace the sodium reduction
189 process. Triglycerides need to be refined to remove free fatty acids, phosphatides, sterols, and oxidation
190 products as well as debris. This step is followed by esterification to produce fatty acid methyl esters and/or
191 hydrogenolysis of methyl esters or fatty acids in the presence of a catalyst at high pressure and
192 temperature to produce methanol and fatty alcohol. The final products are refined or distilled (Kreutzer,
193 1984).

194 In the Lurgi process fatty acids are first converted to wax esters and then hydrogenated over a fixed bed
195 reactor. This differs from the Davy process since there is no conversion to methyl esters and fatty alcohols
196 come directly from wax esters. The Lurgi process has been in commercial use since 2004. The Davy Process
197 is used primarily for detergent alcohols with greater than 12 carbons; however, it can also be used for the
198 production of plasticizer alcohols containing between 6 and 12 carbons. The Davy process provides an
199 improved process for production of fatty alcohols by hydrogenation of lower alkyl esters, particularly
200 methyl esters of fatty acids derived from natural triglycerides under conditions that minimize formation of
201 byproduct alkanes and ethers followed by refining of the resulting ester containing product (Wilmott et al.,
202 1992). Many production plants throughout the world have been licensed to produce fatty alcohols using
203 the Davy process (Fig. 1).

204 Fatty alcohols are also produced synthetically from petroleum. Alkylaluminum derivatives are produced
205 by adding hydrogen and ethylene to an aluminum slurry. Alkylaluminum reacts with ethylene to increase
206 carbon chain length. Higher trialkylaluminum species produced by reacting ethylene with alkylaluminums
207 under pressure at about 120°C can be further reacted with ethylene at higher temperatures to give straight
208 chain alcohols with up to 22 carbons (alfene process). Reaction of the higher trialkylaluminums with air
209 and sulfuric acid yields higher n-alcohols: alfol process (Atwood, 1963; Continental Oil, 1962; 1973). The
210 choice of catalyst and reaction conditions significantly affect the process (Miller, 1969). For fatty alcohol
211 production, it is difficult to practice an esterification on a continuous basis, thus it is convenient to adopt
212 batch processing.

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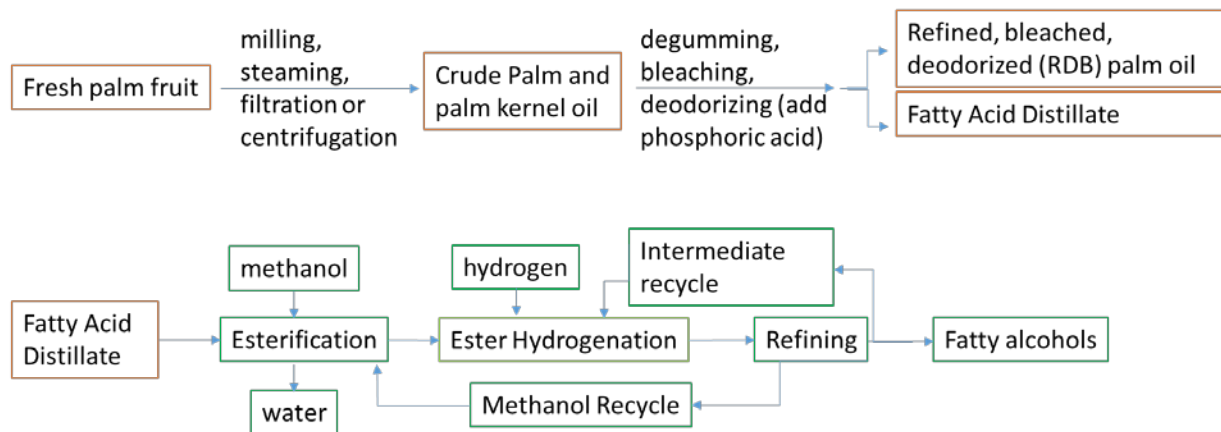


Fig. 1 Davy process for Fatty Alcohols production from olefins (Palm Oil)

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Evaluation Question #3: Discuss whether the petitioned substance is formulated or manufactured by a chemical process, or created by naturally occurring biological processes (7 U.S.C. § 6502 (21)).

219 The present world capacity of plant derived and petroleum derived fatty alcohols is greater than two
220 million metric tons/year. Much of this production goes in to making detergents or plastics. Petroleum
221 derived fatty alcohols production is estimated to be just 23% of total global capacity, whereas plant derived
222 fatty alcohols production is currently 77% of total global capacity (Oleoline, 2015). In the USA the bulk of
223 fatty alcohols is of petrochemical origin, whereas in Europe more than 60% of the total volume is made
224 from natural fats and oils (Kreutzer, 1984).

225 Fatty alcohols occur in free and esterified form in many animal and vegetable waxes. Although these waxes
226 are normally composed of long carbon chains, 1-octanol and 1-decanol have also been found naturally in
227 various food items (Mahadevan, 1978). The aliphatic alcohols are natural components of apples and
228 oranges, and have been reported as a component of edible seeds, oils and fermented beverages (EPA,
229 2007). Aliphatic hydrocarbons can be converted to fatty alcohols by many organisms including bacteria
230 with enzymes in the family of fatty acyl-CoA reductases (Roessler et al., 2014). Because fatty alcohols have
231 important industrial uses as medicines, cosmetics, skin care products, detergents, fuels and plasticizers in
232 addition to their role in tobacco production, extensive research to develop fermentation systems producing
233 fatty alcohols has been undertaken over the past few years with the intention of industrial production in
234 the near future (McCoy et al., 2010; Kallio et al., 2014). Naturally found strains of bacteria including
235 *Escherichia coli*, *Salmonella spp.*, *Klebsiella spp.*, and *Enterobacter spp.* are known to excrete 1-octanol, 1-decanol
236 and 1-dodecanol. Several, *E. coli* strains being examined for commercial scale production of 1-octanol and
237 1-decanol respectively produced as much as 508 and 740 nanograms/milliliter of culture (Hamilton-Kemp
238 et al., 2005).

239 Mutations introduced into *Escherichia coli* strains have resulted in a number of commercially viable
240 fermentation approaches to produce n-alcohols (Dellomonaco et al., 2011). By favoring chain elongation in
241 an anaerobic bacterial growth rescue cycle, enzymes for the production of n-hexanol and n-octanol can be
242 enriched increasing the production of fatty alcohols (Machado et al., 2012). High levels of fatty alcohols (>
243 350 milligrams/liter of culture) can be produced in *E. coli* from a class of enzymes called carboxylic acid
244 reductases (CAR). These enzymes catalyze the reduction of aromatic and short chain carboxylic acids to
245 their respective aldehydes in the presence of adenosine triphosphate (ATP) and nicotinamide adenine
246 dinucleotide phosphate (Akhtar et al., 2013). Mutated *E. coli* strains that produce upwards of 6.33 grams of
247 fatty alcohol per liter of culture support future alternative industrial fermentation methods for the
248 production of fatty alcohols (Liu et al., 2016).

Evaluation Question #4: Describe the persistence or concentration of the petitioned substance and/or its by-products in the environment (7 U.S.C. § 6518 (m) (2)).

251 Alcohols with chain lengths up to C₁₈ including hexanol, octanol, decanol, dodecanol, tetradecanol,
252 hexadecanol and octadecanol are readily biodegradable within ten days. Production plant effluents often
253 appear to only contain naturally occurring products, in spite of fatty alcohols introduced into their

254 influents. This is because microbial action has rapidly altered the influent alcohols to naturally occurring
 255 products in the effluent (Mudge et al., 2014; Stahl and Pessen, 1953). Fatty alcohols are relatively stable
 256 with respect to abiotic degradation in water. Photo-oxidation in aqueous systems is not significant.
 257 Alcohols have no hydrolysable groups and are therefore not susceptible to hydrolysis. Fatty alcohol
 258 oxidation is expected under normal environmental conditions. The fatty alcohols are susceptible to
 259 atmospheric degradation by hydroxyl radicals, with half-lives ranging between approximately 10-30 hours.
 260 Longer chain lengths have shorter estimated half-lives within this range (OECD, 2006). Fatty alcohols are
 261 used in the manufacture of surfactants for detergents and personal care products. These products are
 262 mostly disposed of down the drain at a rate of about 185,000 metric tons per year. Most use is as laundry
 263 detergent totaling about 532,000 metric tons per year. Fatty alcohols used for detergent production are
 264 mostly sourced from petroleum products in the US. By comparison, the contribution of fatty alcohols to the
 265 environment from tobacco topping and suckering is very small (Mudge and DeLeo, 2014).

266 **Evaluation Question #5: Describe the toxicity and mode of action of the substance and of its**
 267 **breakdown products and any contaminants. Describe the persistence and areas of concentration in the**
 268 **environment of the substance and its breakdown products (7 U.S.C. § 6518 (m) (2)).**

269 The octanol-water partition (Log K_{ow}) coefficient is a laboratory measurement of the hydrophilic-lipophilic
 270 balance of a substance. It measures the tendency of a substance in the environment to prefer an organic or
 271 oily phase, e.g., fish (fat) and muscle or sediment, rather than an aqueous phase. The compound is first
 272 dissolved in water to a known concentration and placed in an octanol-water system and allowed to
 273 equilibrate. The concentration in the aqueous phase is re-determined using UV-visible light spectroscopy
 274 (Dalyrimple, 2005). The K_{ow} represents the membrane lipid-water barrier because octanol ($C_7H_{15}CH_2OH$)
 275 represents lipids in living organisms. Lipids and octanol have a similar carbon to oxygen ratio. Thus, it is
 276 used as an indicator of bioconcentration and bioaccumulation in the environment. The density, solubility
 277 and K_{ow} values for some fatty alcohols are given in Table 4. K_{ow} values are generally low for the C_6 - C_{12}
 278 alcohols and increase with increasing molecular weight.

Name	Carbon Number	Molecular Weight, daltons	Density(d^{20}) grams/ml	Solubility @25°C- grams/liter	Log K_{ow}
Octanol	8	130.2	0.827	0.5	3.15*
Nonanol	9	144.3	0.828	0.1	3.77
Decanol	10	158.3	0.830	0.04	4.57
Undecanol	11	172.3	0.832	0.008	4.72
Dodecanol	12	186.3	0.831@24°C	0.00019	5.36

from Mudge, 2005; Fisk et al., 2009

*octanol-water partition coefficient used to measure K_{ow}

279

280 **Evaluation Question #6: Describe any environmental contamination that could result from the**
 281 **petitioned substance's manufacture, use, misuse, or disposal (7 U.S.C. § 6518 (m) (3)).**

282 The Safer Chemical Ingredients List (SCIL) lists chemical ingredients that EPA's Safer Choice Program has
 283 evaluated and determined to be safer than traditional chemical ingredients. This list was designed to help
 284 manufacturers find safer chemical alternatives to meet the criteria of the Safer Choice Program. Safer
 285 Choice decides to include a chemical on the SCIL based on the hazard information from a broad set of
 286 resources, including the identification and evaluation of all available toxicological and environmental fate
 287 data. According to the Safer Choice determination of the EPA, 1-decanol, 1-octanol, 1-dodecanol and the
 288 C_6 - C_{12} alcohols are expected to be of low concern based on experimental and modeled data.

289 Linear fatty alcohols in general are easily biodegradable. The solubility of fatty alcohols in water decreases
290 with an increasing C-chain length. Fatty alcohols possess only moderate acute toxicity for aquatic
291 organisms. In general, in their range of water solubility no toxic effects are observed. However, a number
292 of studies were performed with concentrations that are considerably above the water solubility. Available
293 data for fatty alcohols chronic toxicity do not indicate a special toxicological potential (Condea, 2000).

294 Fatty alcohols are recognized as High Production Volume (HPV) chemicals. Global production volume is
295 estimated at over 1.9 million metric tons. Linear to slightly branched log chain alcohols ranging from 6 to
296 22 alkyl carbons (C) biodegrade exceptionally rapidly in the environment (half-lives on the order of
297 minutes); however, due to continuous use and distribution to waste water treatment systems, partitioning
298 properties, biodegradation of alcohol based surfactants and natural alcohol sources, linear chain alcohols
299 are universally detected in waste water effluents. A large fraction of environmentally detected alcohols are
300 naturally derived alcohol from animal, plant and microbiologically mediated biotransformations. The fatty
301 alcohols from both natural and manufactured sources represent a low risk for environmental
302 contamination (Belanger et al., 2009).

303 C₆-C₁₂ fatty alcohols are likely to volatilize quickly, however, longer chain alcohols (C>12) reaching water
304 supplies are not expected to be hydrolytically degraded (EPA, 2007). The shorter chain fatty alcohols
305 (C<12) are degraded by oxidation and hydrolysis (Patterson et al., 1970). In the atmosphere all C-H
306 containing organic substances react with photochemically generated hydroxyl radicals. The half live for
307 photodegradation of the fatty alcohols varies between 12 and 30 hours (measured for 1-hexanol). Fatty
308 alcohols are biodegradable, and those above C₁₁ may be considered potentially bioaccumulative. Alcohols
309 act by non-polar narcosis. Any toxicity produced by the fatty alcohols with chain lengths less than 12 is
310 considered sub-acute with a fifty percent effective concentration (EC₅₀) ranging from 2.0 to 25
311 milligrams/liter (Fisk et al., 2009). The category comprises a homologous series of linear and essentially
312 linear C₆- C₂₂ alcohols. In addition catalysts such as sulfuric acid present an environmental issue (Condea,
313 2000).

314 Increasing carbon chain length leads to a predictable pattern in physico-chemical properties that drives a
315 distinct range of fate behaviors in the environment. Fatty alcohols all have the same mode of
316 ecotoxicological action. In addition, they are all rapidly biodegradable especially at environmentally
317 relevant concentrations. Alcohols are metabolized/bio-transformed in living organisms suggesting that
318 bioaccumulation potentials based on octanol-water partition coefficients are likely to be overestimated.
319 Measured biological concentration factor (BCF) data on fatty alcohols supports the concept that the
320 bioaccumulation potential of these substances will be lower than estimated from log K_{ow}. 1-hexanol and 1-
321 octanol present a hazard for the environment (acute toxicity to fish, daphnids and algae in the range 1 - 100
322 mg/l). However, both of these substances are readily biodegradable. 1-decanol and 1-undecanol present a
323 greater hazard for the environment (high acute toxicity to fish, daphnids and algae, in the range 0.1 - 1
324 mg/l, and/or high chronic toxicity). The substances in this subgroup biodegrade rapidly and
325 environmental monitoring data from seven countries indicates exposures to the environment is anticipated
326 to be low (OECD, 2006).

327 Available toxicity data indicate that aliphatic alcohols are “practically non-toxic” to honey bees (acute
328 contact LD50 > 25 µg/bee). However, given that aliphatic alcohols can be used as Lepidopteran sex
329 inhibitors, there is a potential for sublethal (e.g., reproductive) effects on non-target Lepidopterans, such as
330 butterflies. This potential effect cannot be quantified at this time (EPA, 2007).

331 **Evaluation Question #7: Describe any known chemical interactions between the petitioned substance**
332 **and other substances used in organic crop or livestock production or handling. Describe any**
333 **environmental or human health effects from these chemical interactions (7 U.S.C. § 6518 (m) (1)).**

334 Manual topping and suckering is labor intensive, but does not involve the use of any chemical substance.
335 Mineral oil, cooking oil or paraffin oil are currently the only topping and suckering substances used by
336 organic crop producers (Little et al., 2008). Fatty alcohols are used independently of other topping and
337 suckering chemicals. There is no known interaction between the fatty alcohols that might be used for
338 topping and suckering (Calvert, 1953).

339 **Evaluation Question #8: Describe any effects of the petitioned substance on biological or chemical**
340 **interactions in the agro-ecosystem, including physiological effects on soil organisms (including the salt**
341 **index and solubility of the soil), crops, and livestock (7 U.S.C. § 6518 (m) (5)).**

342 When fatty alcohols are applied to tobacco plants for suckering with a surfactant such as tween 20, an
343 average residue of 1.6 parts per million (ppm) of the applied fatty alcohols and 1.0 ppm of the surfactant
344 remain on the cured leaves. Over 7000 ppm of naturally occurring fatty alcohols are also present in and on
345 the cured leaves (Tso et al., 1975). Fatty alcohols induce a low incidence of polynucleate root tip cells or
346 root tip cells with fragmented nuclei (Tso and Burk, 1969). The fatty alcohols are produced naturally, in all
347 living organisms, from bacteria to man, and thus, are widely present throughout the natural world. In any
348 agro-ecosystem, fatty alcohols will be present from natural sources. The introduction of C₆-C₁₂ fatty
349 alcohols for topping and suckering may produce short term toxicity to many organisms in the range of 1-
350 100 milligrams/liter, however; because the application rate is intermittent and biodegradability and
351 removal rate are high for this substance no readily observable effects occur in the agroecosystem (OECD,
352 2006).

353 **Evaluation Question #9: Discuss and summarize findings on whether the use of the petitioned**
354 **substance may be harmful to the environment (7 U.S.C. § 6517 (c) (1) (A) (i) and 7 U.S.C. § 6517 (c) (2) (A)**
355 **(i)).**

356 The fatty alcohols are of ecological interest because of their widespread use as surfactants, detergents and
357 personal care products. They are “down the drain products.” In addition fatty alcohols are chemicals that
358 naturally occur in all plants and animals. In general, fatty alcohols with carbon chains longer than C₁₂ are
359 found in the environment. Industrially derived fatty alcohols constitute less than 1% of the total amount of
360 fatty alcohols present in the environment, based primarily on stable radioisotope labelling studies (Mudge
361 et al., 2014).

362 Fatty alcohol products have been assessed for their chronic ecotoxicity in North America and Europe
363 (Belanger et al., 2006). Statistical modelling of distributions and toxicities to algae, fish, daphnids and
364 various controlled experimental systems has been performed and evaluated with the results refined to
365 toxic units based model. The model indicates that the overall fatty alcohol products as a result of their high
366 level of biodegradability and low toxic units, 0.015 to 0.212, were considered a very low risk for
367 environmental contamination (Sanderson et al., 2013). Fatty alcohols are known for their high level of
368 biodegradability in the environments. Their derivative products are additionally designed to rapidly
369 degrade after use (Atkinson et al., 2009). Fatty alcohols are not considered endocrine disrupters.

370 Long chain alcohols (LCOH) with a carbon chain length range of C₆-C₂₂ covering 30 substances, and 41.5
371 million metric tons/year consumed globally, were evaluated under the Organization for Economic
372 Cooperation and Development (OECD) high production volume chemicals program in 2006. The main
373 findings of the assessment include: (1) no unacceptable human or environmental risks were identified; (2)
374 these materials are rapidly and readily biodegradable; (3) a parabolic relationship was demonstrated
375 between carbon chain length and acute and chronic aquatic toxicity; (4) category-specific (quantitative)
376 structure-activity relationships were developed enabling prediction of properties across the entire category;
377 (5) LCOH occur naturally in the environment in an equilibrium between synthesis and degradation; (6)
378 industry coming together and sharing resources results in minimizing the need for additional animal tests,
379 produces cost savings, and increases scientific quality of the assessment (Sanderson et al., 2008). Daphnia, a
380 standardized environmental test crustacean, is effected by 1-decanol at a concentration of approximately 1
381 milligram per liter (Schafers et al., 2009).

382 Fatty alcohols have a moderate tendency to bind to soils. The portion of applied chemical binding to the
383 soil, rather than volatilizing, will be subject to biodegradation, with estimated half-lives for 1-octanol and 1-
384 decanol of 2.3 days. The portion of applied chemical that does volatilize is estimated to degrade in the air
385 by reaction with hydroxyl radicals with half-lives of about 10 hours (EPA, 2007).

386 **Evaluation Question #10: Describe and summarize any reported effects upon human health from use of**
387 **the petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (i), 7 U.S.C. § 6517 (c) (2) (A) (ii) and 7 U.S.C. § 6518**
388 **(m) (4)).**

389 Toxicity data for the aliphatic alcohols consisting of acute toxicity, irritation, and sensitization studies,
390 developmental rat (oral and inhalation) toxicity studies and a 90-day rat (dermal) study were evaluated for

391 the Environmental Protection Agency (EPA) health risk determination. The available mutagenicity studies
392 included the Ames, micronucleus, and gene mutation assays. Acute inhalation studies with the rat resulted
393 in estimates of the median lethal dose (LD₅₀) above the limit concentration of 2 milligrams/Liter. However,
394 eye irritation studies resulted in severe and sometimes non-reversible eye irritation. Dermal irritation
395 studies revealed slight to moderate irritation in rabbits, and the aliphatic alcohols generally did not
396 produce sensitization in tests with guinea pigs. There is no evidence to suggest that the aliphatic alcohols
397 cause increased susceptibility in infants and children. Based on the results of the available studies, no
398 endpoints of toxicological concern have been identified for human health risk assessment purposes. The
399 EPA concluded that there are no human health risks of concern for aliphatic alcohols. Currently, there is no
400 known mode of toxicological action for the aliphatic alcohols. Based on the low hazard concern via the oral,
401 dermal, and inhalation routes of exposure, a quantitative risk assessment for the aliphatic alcohols was not
402 found necessary (EPA, 2007).

403 EPA is updating the Restricted Entry Interval (REI) and Personal Protective Equipment (PPE) requirements
404 for aliphatic alcohols. 1-Decanol, which is a component of all the tobacco sucker control products in this
405 case, is an acute Toxicity Category I eye irritant; therefore, pursuant to the Worker Protection Standard
406 (WPS) and according to the EPA Office of Pollution Prevention and Toxics (OPPTS) Label Review Manual
407 3rd Edition, products with agricultural uses must require a 48 hour REI and the following personal
408 protective equipment (PPE) for early entry: coveralls, chemical-resistant gloves made of any water proof
409 material, shoes plus socks, and protective eyewear (EPA, 2007).

410 Fatty alcohol products used for tobacco topping and suckering are of not very toxic following acute and
411 repeated exposures. Although, skin and eye irritation are commonly observed. The mammalian
412 metabolism of fatty alcohols used in tobacco topping and suckering is highly efficient. (Veenstra et al.,
413 2009; OECD, 2006). Aliphatic alcohols are absorbed by all common routes of exposure, widely distributed
414 within the body and efficiently eliminated. There is a limited potential for retention or bioaccumulation for
415 the parent alcohols and their biotransformation products. Overall, toxicology databases show an inverse
416 relationship between chain length and toxicity. The shorter chain alcohols tend to induce more pronounced
417 effects when compared to materials with a longer chain length. This is illustrated most clearly by the
418 degree of irritation in skin and eye irritation studies in laboratory animal studies. For the aliphatic alcohols
419 in the range C₆ - C₁₁ a potential for skin and eye irritation exists, without concerns for tissue destruction or
420 irreversible changes. Aliphatic alcohols in the range C₁₂ - C₁₆ have a low degree of skin irritation potential.
421 The eye irritation potential for alcohols with a chain length of C₁₂ and above has been shown to be minimal.
422 Aliphatic alcohols have no skin sensitization potential. Repeated exposure to aliphatic alcohols is generally
423 without significant systemic toxicological findings and regarded to be of a low order of toxicity upon
424 repeated exposure. C₆-C₁₂ can induce local irritation at the site of first contact. There is a suggestion of mild
425 change consistent with low-grade liver effects. Typical findings include: slightly increased liver weight, in
426 some cases accompanied by clinical chemical changes but generally without concurrent histopathological
427 effects. Aliphatic alcohols do not have a potential for producing peroxisome proliferation. Central nervous
428 system (CNS) effects were absent upon inhalation or dietary administration, however 1-hexanol and 1-
429 octanol showed a potential for CNS depression upon repeated administration of a bolus dose. Similarly, 1-
430 hexanol and 1-octanol induced respiratory distress upon repeated administration of a bolus dose. Aliphatic
431 alcohols do not have a potential for peripheral neuropathy. Typical no observable adverse effects level
432 (NOAEL) for aliphatic alcohols range from 200 mg/kg/day to 1000 mg/kg/day in the rat upon sub-
433 chronic administration via the diet. There has not been evidence of a carcinogenic potential for aliphatic
434 alcohols. They do not contain structural elements of concern for potential interaction with DNA and have
435 been shown to be without mutagenic activity, primarily on the basis of Ames assays and mouse
436 micronucleus assays (Nelson et al., 1990a; OECD, 2006).

437 On the basis of the lack of adverse findings in the reproductive organs in repeated dose toxicity studies and
438 in screening studies for reproductive effects aliphatic alcohols are considered without a potential for
439 adverse effects on fertility and reproductive toxicity. Similarly, developmental toxicity studies with
440 aliphatic alcohols have confirmed the lack of potential adverse effects on the developing fetus. As a rule
441 aliphatic alcohols are manufactured and processed in established chemical complexes in closed
442 installations; these are usually operated at high temperature and pressure. At these sites standard personal
443 protective equipment is routinely applied to prevent direct skin and eye contact. Generally, aliphatic
444 alcohols are of a low volatility and as a rule engineering controls are available preventing the need for

445 respiratory protection. For non-routine operations involving a break in enclosed systems a higher level of
446 protection is applied. Operations with a potential for significant exposure require a permit to work system
447 and a case-by-case assessment is made for appropriate protective measures. Exposure through the use of
448 products in industry and commerce is mitigated by applying measures aimed to prevent direct skin and
449 eye contact by following the recommendations in the material safety data sheet (MSDS). Aliphatic alcohols
450 are formulated in consumer laundry, cleaning and personal care products. Product labels reflect the hazard
451 potential of the chemical ingredients in these products and include first aid instructions in case of non-
452 intentional exposure (Nelson et al., 1990b; OECD, 2006).

453 **Evaluation Question #11: Describe all natural (non-synthetic) substances or products which may be**
454 **used in place of a petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (ii)). Provide a list of allowed**
455 **substances that may be used in place of the petitioned substance (7 U.S.C. § 6518 (m) (6)).**

456 Tobacco is apically dominant. The growing point via plant hormones exerts an influence throughout the
457 vegetative and most of the reproductive phases, preventing the development of the axillary buds. Topping
458 causes the axillary buds of the upper leaves to develop rapidly into suckers that must be removed at
459 regular intervals until the final harvest. The suckers provide a harbor for insects and plant diseases. The
460 use of a chemical for suckering and/or for topping results in a saving of labor, reduced infestations of
461 insect pests, reduced spread of plant diseases, for example mosaic. There would also be less damage to the
462 crop as a result of less travel through it. Indoleacetic acid at 10,000 ppm applied in lanolin will prevent
463 sucker growth. Mineral oil daubed on the stem after topping also prevents sucker growth (Calvert, 1953).

464 Ten percent neem oil, 20% mohua oil, 25% groundnut oil and mineral oil applied to each plant after
465 topping were evaluated against hand topping and suckering. All of these treatments improved sucker
466 control and yield (Bangaryya et al., 1982).

467 Methyl caprate has been used effectively for topping and suckering. Although this fatty acid is an
468 oleochemical, it may also be possible to produce it by fermentation (Tso, 1964).

469 Mineral oil, properly used can produce results in chemical suckering that are similar to hand suckering.
470 However, some mineral oils may be too dangerous to use or must be diluted (emulsified) otherwise the oil
471 may not only remove the suckers, but also burn the leaves. Mineral oil treatment is affected by rain. The
472 use of mineral oil saves labor costs and exposure of personnel to nicotine and potentially green tobacco
473 syndrome. Labor savings can be significant and as much 50% (Salmon, 1959; Wilson et al., 1952).

474 Apical dominance in tobacco can be controlled with plant growth hormones. However the removal of the
475 tobacco inflorescence, topping acts as an immediate stimulus for the growth of suckers. The use of mineral
476 and vegetable oil for suckering has the potential to cause or exacerbate soft rot. The cause may be too much
477 oil and it running down the plant to contact roots. Highly refined mineral oil with very low aromatic
478 content should be used. Liquid paraffin, white oil and light medicinal oil have been successfully used for
479 suckering. Although it takes about twice as long to apply the oil as it does to hand sucker, subsequent
480 manual suckering sessions are eliminate saving time and labor (Calvert, 1956b).

481 .

482 **Evaluation Question #12: Describe any alternative practices that would make the use of the petitioned**
483 **substance unnecessary (7 U.S.C. § 6518 (m) (6)).**

484 When the tobacco crop is about half-grown, flower buds begin to appear. These flower heads are removed
485 or "topped" to prevent seed formation, forcing the plant to focus on leaf production. The result is larger,
486 thicker, darker leaves that mature more uniformly and contain more nicotine. Topping may be done by
487 hand or with special machines that cut the flower heads and sacrifice a few leaves. Topping requires two or
488 three trips over the field to catch all the plants. Topping of plants also stimulates the growth of secondary
489 stems from the base and/or leaf axils. These "suckers" must also be removed to assure uniformity and
490 quality. While chemicals are available to suppress suckering, these may not be allowable under organic
491 certification standards. The alternative is removal by hand every seven to ten days. Suckering is one of the
492 most labor-intensive activities in tobacco production, as many plants sucker two or three times before
493 harvest (Kuepper and Thomas, 2008).

494 The aim of sucker control is to focus the plant's energy into filling the leaves rather than growing the
495 flower. Because tobacco sells by weight, heavier leaves are favored economically. In organic tobacco

496 production, early topping to improve yield and quality is usually done by hand. Suckers can be removed
497 by hand as well as stunted by carefully applying approved soybean oil or mineral oil to the top of the
498 plant. Topping and suckering are the most time consuming tasks associated with growing organic tobacco,
499 and may be necessary every week for 10 weeks. It can take one person per acre per day to do the job. Plants
500 may not be permitted to flower, instead, they are topped at 15 leaves. In organic tobacco production,
501 controlling insects without insecticides and weeds without herbicides is not as big a big problem as
502 controlling suckers. Application of mineral or cooking oil to prevent the suckers from coming back has to
503 be done mostly by hand. Initial topping and suckering is done by hand. Vegetable oil or mineral oil is
504 poured from gallon jugs over each plant, allowing the oil to run into each leaf axil to the way to the bottom
505 of the plant. Topping early keeps the aphid off tobacco plants. Soybean cooking oil and mineral oil applied
506 to the heads of plants prevents flowering. Eighteen hours can be spent per acre removing suckers (Little et
507 al., 2008). There are currently 314 certified organic tobacco production operations. All of them are located
508 in the United States of America (Organic Integrity Database).

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