

Vitamins B₁, C and E

Crops

Identification of Petitioned Substance

Chemical Names:

Vitamin B₁: 2-[3-[4-Amino-2-methyl-pyrimidin-5-yl)methyl]-4-methyl-thiazol-5-yl] ethanol

Vitamin C: (R)-3,4-dihydroxy-5-((S)-1,2-dihydroxyethyl)furan-2(5H)-one

Vitamin E: (2R)-2,5,7,8-Tetramethyl-2-[(4R,8R)-(4,8,12-trimethyltridecyl)]-6-chromanol

CAS Numbers:

67-03-8 (Thiamine hydrochloride)

50-81-7 (L(+)-Ascorbic acid)

59-02-9 ((+)- α -Tocopherol)

Other Codes:

200-641-8 (Thiamine hydrochloride)

200-066-2 (L(+)-Ascorbic acid)

200-412-2 ((+)- α -Tocopherol)

Other Name:

Thiamine (Vitamin B₁)

Ascorbic Acid (Vitamin C)

Tocopherols (Vitamin E)

Trade Names:

Dyna-Gro, Rhizo Gel (B₁ products); Thiamine hydrochloride, L-Ascorbic acid, α -Tocopherol

Summary of Petitioned Use

The National Organic Program (NOP) final rule currently allows the use of vitamins B₁, C and E as plant or soil amendments in organic crop production under 7 CFR §205.601(j)(8). Synthetic forms of vitamins – including vitamins B₁, C and E – are also permitted in other areas of organic production and handling. In addition to crop applications, nutrient vitamins are also allowed as feed additives in organic livestock production under 7 CFR §205.603(d)(3) in amounts needed for nutrition and health maintenance (7 CFR 205.237). Similarly, synthetic sources of vitamins may also be incorporated into processed products labeled as “organic” or “made with organic (specified ingredients or food group(s))” (7 CFR 205.605(b)). This report provides targeted technical information on vitamins B₁, C and E for the National Organic Standards Board’s review of these substances under the sunset process.

Characterization of Petitioned Substance

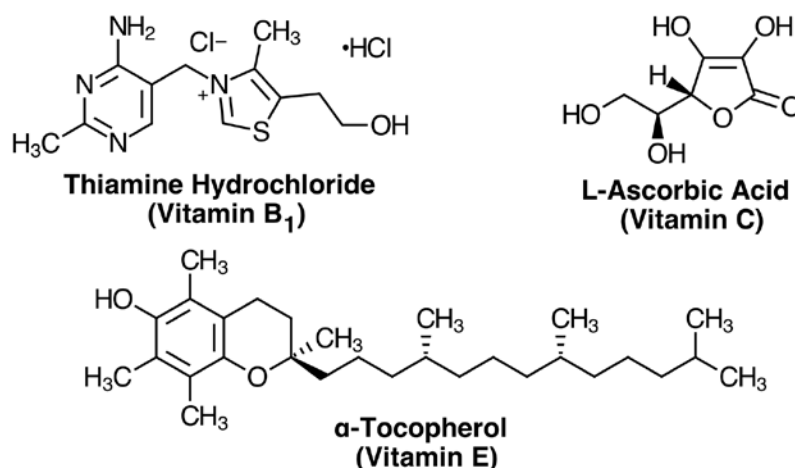
Composition of the Substance:

Vitamins B₁, C and E are naturally occurring compounds synthesized in small amounts by various plants and microorganisms, including bacteria and fungi. At the molecular level, water-soluble vitamins in this group (i.e., vitamins B₁ and C) consist of carbon and hydrogen as well as several electronegative atoms, including nitrogen, oxygen, sulfur and chlorine. The abundance of polar functionality – amino groups, hydroxyl groups, and positively or negatively charged atoms – imparts enhanced water solubility to vitamins B₁ and C. Vitamin E, on the other hand, is composed predominantly of non-polar carbon-carbon and carbon-hydrogen bonds, and contains only one polar functionality (i.e., hydroxyl group) on the substituted benzopyran subunit. Vitamin E is classified as a fat-soluble vitamin. Figure 1 below provides two-dimension representations of the molecular structures for vitamins B₁, C and E.

Source or Origin of the Substance:

Vitamins can be extracted from foods or synthesized by chemical or biofermentation processes. Regarding the former method, certain vitamins can be obtained from natural dietary sources in varying quantities. For example, vitamin C (ascorbic acid) is a major nutritional component of citrus fruits and green leafy vegetables. Vitamins B₁ and C are produced on an industrial scale using chemical synthesis, while commercially available sources of vitamin E are obtained through a combination of extraction and chemical

49 synthesis (Festel, 2005). The available patent literature indicates tremendous growth in the development of
50 fermentative methods for the production of vitamin compounds utilizing genetically modified
51 microorganisms (GMMs) over the past decade. See the response to Evaluation Question #2 for detailed
52 information regarding currently utilized industrial production techniques and an analysis of trends in the
53 application of GMMs in the synthesis of vitamins B₁, C and E.



54

55 **Figure 1. Water-soluble vitamin B₁ and C contain polar functionalities, while fat-soluble vitamin E**
56 **contains largely non-polar chemical bonds.**

57 Properties of the Substance:

58 As a result of the structural diversity among the vitamin compounds, there is great variability in the
59 physical and chemical properties of vitamins as a chemical class. Vitamins are organic (i.e., carbon-based)
60 compounds and are typically grouped depending on their solubility in water vs. organic solvents. The
61 more hydrophilic vitamin compounds (vitamin B₁ and C) tend to have multiple polar functionalities, such
62 as hydroxyl groups, amino groups, carboxylic acids, alkoxy groups, and/or salts of carboxylic acids and
63 amines. Due to their enhanced aqueous solubility, vitamin B₁ and C molecules not absorbed or metabolized
64 by the organisms are rapidly excreted in biological fluids. Alternatively, more lipophilic (fat-soluble)
65 vitamins such as vitamin E are primarily composed of aliphatic carbon frameworks and may be stored in
66 fatty tissues upon excessive consumption of the vitamin. As a class of substances, vitamins have relatively
67 low vapor pressures (HSDB, 2010a; HSDB, 2010b). Details regarding the physical and chemical properties
68 for vitamins included in this review (i.e., vitamins B₁, C and E) are provided in the following subsections.

69 *Vitamin B₁*

70 Vitamin B₁ (thiamine) and thiamine hydrochloride, a commonly used industrial form of vitamin B₁ are
71 colorless solids with melting points of 164 and 250 °C (HSDB, 2010a; ChemicalBook, 2010a). One gram of
72 thiamine dissolves in approximately 1 mL water, 18 mL glycerol, 100 mL 95% alcohol, or 315 mL absolute
73 alcohol (ethanol); thiamine is practically insoluble in non-polar organic solvents such as diethyl ether,
74 benzene, hexane and chloroform. The pH of a 1% weight/volume solution of thiamine in water is 3.13
75 (HSDB, 2010a). Chemical forms of vitamin B₁ are generally light sensitive and hygroscopic (i.e., readily
76 absorb moisture from the atmosphere) (ChemicalBook, 2010a).

77 *Vitamin C*

78 Pure vitamin C (L-ascorbic acid) is a colorless crystalline powder or solid at room temperature. Aqueous
79 solutions of vitamin C have a pH of 1.0–2.5 at 176 g/L at 25 °C. The melting point/range of pure L-ascorbic
80 acid is 190–194 °C. L-ascorbic acid is highly soluble in water (solubility = 176 g/L at 20 °C). In addition, L-
81 ascorbic acid exhibits air and light sensitivity and, as an antioxidant, it acts as a strong reducing agent with
82 some organic compounds (Sigma Aldrich, 2015; Fisher Scientific, 2012).

83 *Vitamin E*

84 The most biologically active form of vitamin E is α -tocopherol. It exists as a yellow-brown viscous oil with
85 a melting point/range of 200–220 °C and a density of 0.95 g/mL at 20 °C. As a fat-soluble vitamin, all forms
86 of vitamin E are insoluble in water and soluble in many non-polar organic solvents. Due to its antioxidant
87 properties, vitamin E may also react violently with oxidizing agents. Combustion of vitamin E may lead to
88 the production of carbon oxides (Sigma Aldrich, 2014; ChemicalBook, 2010b).

89 **Specific Uses of the Substance:**

90 Synthetic forms of vitamins B₁, C and E enjoy a variety of uses ranging from the stimulation of crop growth
91 and plant protection to supplementation in livestock feeds and human dietary supplements. This section
92 summarizes the uses of these specific vitamin compounds, with a focus on their application in organic and
93 conventional crop production.

94 *Root Stimulation Using Vitamin B₁*

95 Vitamin B₁ is included in several commercially available root stimulator products to help nursery-grown
96 mature plants, seedlings and cuttings become established when transplanted to native soil (Schalau, 2010).
97 According to the Organic Materials Review Institute (OMRI) Generic Materials List, growth regulators for
98 plants include non-synthetic plant hormones such as gibberellic acid, indole acetic acid (IAA) and
99 cytokinins. Synthetic forms of vitamin B₁ are also approved for use in organic crop production as a growth
100 regulator for plants. Organically approved growth stimulator products containing vitamin B₁ were not
101 identified on the OMRI product list (OMRI, 2013).

102 Nursery operators and farmers commonly use vitamin B₁ root stimulator products to prevent transplant
103 shock and stimulate the growth of new roots when planting trees, shrubs, roses and other plants (Cox,
104 2010). Many conventional products marketed as root stimulators list vitamin B₁ as an essential ingredient;
105 however, these products also contain auxins – such as the synthetic plant hormone indole butyric acid
106 (IBA) and naphthylacetic acid (NAA) – as well as various fertilizer compounds (Schalau, 2010). The bulk of
107 the available scientific data suggests that IBA and NAA contribute to root regeneration of transplanted
108 trees by suppressing crown growth to effectively redirect resources to developing roots. Conversely, the
109 root growth claims associated with vitamin B₁ are largely unsubstantiated and generally based on tissue
110 culture research in which plant tissues are propagated using sterile conditions and artificial growth media
111 (Schalau, 2010; Kontaxis & Cox, 1984). According to Cox (2010), “several studies using intact mums, apple
112 trees, orange trees, pine, tomato, beans, pepper, corn, pear, watermelon and squash have failed to
113 demonstrate that vitamin B₁ treatments provide any type of growth response.” For example, Kontaxis &
114 Cox (1984) transplanted 19-day old seedlings of tomato, green pepper, squash, watermelon, sweet corn,
115 snap bean and pole bean, applying thiamine hydrochloride solutions to the soil of treatment groups at
116 planting and once several days later. No difference was observed between vitamin B₁-treated and control
117 plants in terms of vigor, size, color or root development at 18 to 29 days after first treatment.

118 *Plant Protection and Growth Stimulation Using Vitamins C and E*

119 Practical information regarding the use of vitamins C and E in modern crop production from agricultural
120 extension service websites and related resources are generally unavailable. Therefore, specific use
121 information for these two synthetic substances in crop production is based largely on information derived
122 from the peer-reviewed scientific literature.

123 The antioxidant qualities of vitamins E and C have been successfully employed to promote growth/yield
124 and protect plants from oxidative stress due to salinity. El-Tohamy & El-Greadly (2007) found that foliar
125 applications of vitamin E (0.1 mL/L) to 20-day-old bean plants resulted in statistically significant increases
126 in growth, yield, bean pod quality, natural plant growth hormone levels, as well as chlorophyll and
127 mineral (nitrogen, phosphorus and potassium) contents in leaves. The antioxidant properties of vitamin E
128 have also been utilized in the treatment of Jonagold apple trees, which resulted in a 6-fold increase in the
129 fruits within 48 hours following application to leaf discs (Noga & Schmitz, 2000). Hussein et al. (2007)
130 demonstrated that foliar application of vitamin E (200 ppm) to cowpea plants under salinity stress
131 significantly improve plant health parameters, including plant height and dry weight. Vitamin E produced
132 within the plant (i.e., endogenous vitamin E) has also been found to play a key role in low-temperature

133 adaptation and nutrient transport in the phloem (Maeda, 2006). Research conducted using live apple
134 seedlings and trees found that foliar applications of vitamin E in glycerol and water (0.25%) protected the
135 leaves of apple seedlings as well as flowers of orchard trees against freezing injury (Albrecht, 2004).

136 Recent research indicates that application of vitamin C to maize and kidney bean plants are protective of
137 changes in growth and metabolic activity associated with salt-induced stress (Hassanein, 2009; Salama,
138 2014). According to Hassanein et al. (2009), application of vitamin C as a shoot spray or grain soaking “did
139 not only alleviate the inhibitory effect of salinity stress on the biosynthesis of photosynthetic pigments, but
140 also induced a significant stimulatory effect greater than observed in the corresponding controls.” The
141 potential antioxidant mechanisms of vitamins C and E associated with plant protection and enhanced
142 growth are described below in “Action of the Substance.” A strong body of evidence for commercially
143 grown plants indicates that vitamin C applied as an aqueous spray solution of ascorbate salts (e.g.,
144 potassium ascorbate) moves into leaf cells and increases the tolerance of these cells to the damaging effects
145 of smog in some plants (Freebairn, 1960). Indeed, a more recent study demonstrated that application of
146 foliar sprays containing vitamin C at 300 mg/L to broad beans minimizes the adverse effects associated
147 with exposure to the oxidant ozone and other air pollutants, such as nitrogen oxides and sulfur dioxide
148 (Ali & Musallam, 2007). Application of foliar sprays containing vitamin B₁₂ (50 ppm), folic acid (50 ppm)
149 and vitamin C (500 ppm) to wheat at 30 and 60 days after planting encouraged plant growth, grain yield
150 and mineral contents in leaves and grains relative to control plants (Mohamed, 2013).

151 Patents have been developed based on the ability of antioxidants such as vitamins C and E to strengthen
152 the defense systems of treated plants. Indeed, one disclosed method involves the application of vitamin C
153 and E to plants as either soil drenches or foliar sprays to increase resistance to pests and pathogens
154 associated with common plant diseases (Norris, 1991). Likewise, Abdel-Kader et al. (2012) used ascorbic
155 acid as a resistance chemical inducer in combination with biological control agents (e.g., *Bacillus subtilis*) to
156 reduce the incidence and severity of foliar plant disease, such as powder/downy mildews of cucumber,
157 cantaloupe and pepper as well as early and late blights of tomato.

158 *Vitamins B₁, C and E as Nutrient Vitamins in Livestock Production and Dietary Supplements*

159 Organically- and conventionally-raised livestock are regularly provided with synthetic forms of water-
160 soluble (B-complex and C) and fat-soluble vitamins to supplement the nutrient loads naturally found in the
161 diet. Vitamins are commonly supplemented by injection (vitamins A, D and E); fortification of grain mixes
162 or silage-based rations to ensure each animal receives some vitamins each day; and free choice
163 supplementation through free choice mineral supplements, protein licks/blocks or in salt/mineral/vitamin
164 mixes (Alberta, 2015). Ruminants such as cattle and sheep are able to generate virtually all of the require B-
165 complex vitamins from the raw materials in their diet; nutrient premixes for ruminants generally contain
166 vitamins A, D and E (Wahlberg & Greiner, 2006). In contrast, swine and poultry must obtain a greater
167 number of vitamins through the diet. Because of this limitation and the nutritional variability of natural
168 feed materials, many livestock producers supplement feed sources with vitamins A, D, E and K as well as
169 specific B-vitamins that may otherwise be deficient in feed (NRC, 1994; NCSU, undated).

170 Human dietary supplements generally contain a combination of essential nutrients, including synthetic
171 vitamins such as thiamine, ascorbic acid and tocopherols. Higher intakes of vitamins can be particularly
172 important for recovery following surgical operations. Indeed, recent scientific research suggests that intake
173 of 500 mg/day of the antioxidant vitamin C is associated with reduced postoperative oxidative stress
174 (Fukushima, 2010). Additionally, food products are commonly fortified with vitamins B₁, C and E and
175 other essential nutrients to facilitate sufficient public consumption of these vital compounds (FDA, 2015).

176 **Approved Legal Uses of the Substance:**

177 Vitamins B₁, C and E are legally allowed for use as feed additives for animal production and supplements
178 for human consumption. This section summarizes the legal uses of these specific vitamin compounds
179 according to relevant federal regulations.

180 *Ingredients in Conventional and Organic Livestock Feed*

181 The U.S. Food and Drug Administration (FDA) enforces provisions of the Federal Food, Drug and
182 Cosmetic Act (FFDCA) associated with additives used in animal feed and food for human consumption.

183 According to the FFDCA, any substance that is added or expected to directly or indirectly become a
 184 component of animal food must be used according to the relevant food additive regulation unless the
 185 substance is generally recognized as safe (GRAS) under 21 CFR 582 and 584 for that use pattern (FDA,
 186 2014a). In addition, substances listed as FDA-approved food additives (21 CFR 570, 571, and 573) may also
 187 be incorporated into animal feeds.

188

189 The following synthetic forms of vitamins B₁, C and E are classified as GRAS by the FDA and therefore are
 190 not subject to additional regulatory oversight (OMRI, 2013):

- 191 • Vitamin B₁ (thiamine hydrochloride) 21 CFR 582.5875
- 192 • Vitamin C (ascorbic acid) 21 CFR 582.5013
- 193 • Vitamin E (α -tocopherol acetate) 21 CFR 582.5892

194 *Human Food Additives and Dietary Supplements*

195 The National Organic Program (NOP) final rule currently allows nutrient vitamins in the organic handling
 196 of food for human consumption under 7 CFR 205.605, synthetic substances allowed as ingredients in or on
 197 processed products labeled as “organic” or “made with organic (specified ingredients or food group(s)).”
 198 Organic handlers must also comply with the FDA Nutritional Quality Guidelines for Foods (21 CFR 104.20)
 199 in the fortification of processed foods. The nutrient profiles are provided below (Table 1). In contrast to its
 200 role in the regulation of drugs and animal feed additives, the FDA does not regulate human dietary
 201 supplements; however, if an unsafe product is marketed, it is the responsibility of the FDA to take any
 202 necessary regulatory action and/or ensure the accuracy of the supplement’s label (FDA, 2014).

203 **Table 1. FDA Nutrition Quality Guidelines for Foods: Vitamins B₁, C and E**

Vitamin	Unit of Measurement	DRV or RDI	Amount per 100 calories
Vitamin C	mg	60	3
Vitamin E	IU	30	1.5
Vitamin B ₁ (thiamine)	mg	1.5	0.08

204 IU = International Unit, unit of activity or potency for vitamins and other substances; mg = milligram
 205 (gram/1,000); DRV = Dietary Reference Values; RDI = Reference (Recommended) Daily Intake

206 **Action of the Substance:**

207 Modes of action have not been clearly elucidated for vitamin B₁, C and E as plant and soil amendments.
 208 Indeed, many argue that soil applications of vitamin B₁ provide no benefit for root growth, and the
 209 proposed plant protection mechanisms for vitamins C and E are based primarily on correlations and
 210 established antioxidant pathways for these compounds in animals and plants. This section summarizes
 211 plausible mechanisms for vitamins B₁, C and E as used in crop production.

212 Root stimulant substances generally work by stimulating the root to produce its own natural rooting
 213 hormones to help increase lateral root growth and root mass (Smith, 2015). However, several sources have
 214 indicated that application of vitamin B₁ to the root systems of whole plants does not actually stimulate root
 215 growth following transplantation (Schalau, 2010; Kontaxis & Cox, 1984). The claims associating vitamin B₁
 216 with enhanced root growth are likely based on tissue culture studies in which vitamin B₁ is commonly
 217 included as an ingredient of the growth media (Schalau, 2010). It is therefore difficult to describe the mode
 218 of action for vitamin B₁ root growth stimulants in the absence of significant *in vivo* results correlating
 219 vitamin B₁ soil applications with enhanced root growth.

220 L-ascorbic acid (vitamin C) serves a predominantly protective role in both plants and animals. The
 221 naturally occurring substance functions as a major oxidation/reduction buffer and as a cofactor for
 222 enzymes involved in regulating photosynthesis, hormone biosynthesis and regenerating other antioxidant
 223 species (Gallie, 2013). It also regulates cell division and growth and is involved in signal transduction.
 224 Foliar sprays and soil drenches of aqueous vitamin C solutions (exogenous sources) can help plants cope
 225 with moderate fluctuations in the endogenous levels of this essential plant nutrient. As a powerful

226 antioxidant, vitamin C defends vulnerable plant tissues against the damaging oxidative effects of ozone by
227 neutralizing free radical species (Burkey, 2003). Researchers have found that plants capable of moving
228 greater quantities of vitamin C into the space surrounding cell walls (apoplasts) have a better change of
229 detoxifying ozone (Burkey, 2003). Scientific results supporting the protective effects of plant and soil
230 amendments containing vitamin C have been disclosed in peer-reviewed journal articles and patents
231 (Freebairn, 1960; Norris, 1991).

232 The stimulatory effects of vitamin E applications on plant height, number of leaves and branches, and yield
233 of various bean species has generally been attributed to the antioxidant properties of vitamin E (El-
234 Tohamy, 2007). While vitamin C is highly water-soluble and generally resides in aqueous compartments
235 outside of cell walls and membranes, vitamin E is a low molecular weight lipophilic (fat-soluble)
236 antioxidant that accumulates within cell membranes (El Bassiouny, 2005). Thus, vitamin E can be
237 characterized as a highly effective antioxidant at the membrane site (Hess, 1993). The concerted actions of
238 fat-soluble vitamin E at the surface of membranes and water-soluble vitamin C within aqueous
239 extracellular spaces provides plant tissues with multiple layers of protection from oxidative damage
240 associated with exposure to ozone, as discussed above for vitamin C. Abiotic stressors – including high-
241 intensity light, salinity, drought and low temperatures – are commonly associated with higher levels of
242 oxidative stress in plants. The tocopherols comprising vitamin E can efficiently quench singlet oxygen (and
243 other reactive oxygen species), scavenge various radicals such as lipid peroxy radicals, and thereby
244 terminate lipid peroxidation chain reactions associated with oxidative stress (Maeda, 2006).

245 **Combinations of the Substance:**

246 Products marketed as root stimulators often list vitamin B₁ in combination with synthetic plants hormones
247 known as auxins as well as various fertilizer compounds (Schalau, 2010). Common auxins include indole
248 butyric acid (IBA) and 1-naphthaleneacetic acid (NAA). For example, Dyna-Grow K-L-N Rooting
249 Concentrate is a vitamin-hormone solution containing vitamin B₁ and both of the synthetic rooting
250 hormones, IBA and NAA, with claims of promoting vigorous root growth in trees, foliage and flowering
251 plants (Hydro Galaxy, 2015). Synthetic plant hormones are not approved for use in organic production;
252 however, naturally occurring auxins such as indole acetic acid (IAA) and other plant growth hormones
253 such as cytokinins and gibberellic acid may be used in combination with vitamin B₁ to regulate plant
254 growth in organic crop production (OMRI, 2013). The prevalent combination in commercially available
255 products of vitamin B₁ with synthetic substances prohibited in organic production suggests that organic
256 operators must obtain technical forms of vitamin B₁ (e.g., thiamine hydrochloride) lacking synthetic
257 hormones for use as a rooting agent.

258 Less is known regarding materials typically applied with vitamins C and E for plant protection in organic
259 or conventional crop production. Albrecht et al. (2004) demonstrated the synergistic effect of mixtures
260 containing the antioxidant α -tocopherol (vitamin E) and the cryoprotector glycerol against freezing injury
261 in the leaves and flowers of apple trees. In addition, Norris (1991) described how co-application of vitamins
262 C and E to the surface of plants confers enhanced resistance to environmental stresses associated with pest
263 insects and pathogenic microorganisms. The defensive response elicited through application of vitamins C
264 and E is systemic in nature; treatment of one portion of a plant initiates a defensive response throughout
265 the plant (Norris, 1991).

266 **Status**

267

268 **Historic Use:**

269 In 1995, the National Organic Standards Board (NOSB) recommended addition of vitamins B₁, C and E to
270 the National List as approved plant and soil amendments in organic crop production (USDA, 1995).
271 Researchers observed a positive correlation between vitamin B₁ application and the rate of root growth in
272 plant tissue culture studies conducted in the late 1930s (Bonner & Greene, 1938). Around this time,
273 naturally occurring plant growth regulators known as auxins were isolated and found to stimulate cell
274 elongation in roots and stem tissue (Chalker-Scott, undated; Hoffman, 2010). Manufacturers have therefore
275 co-formulated plant hormones (e.g., synthetic auxins) and vitamin B₁ in root stimulation products since the

276 late 1940s (Chalker-Scott, undated). No historical information relevant to the current review was identified
277 regarding the use of vitamins C and E in organic or conventional crop production.

278 **Organic Foods Production Act, USDA Final Rule:**

279 Vitamins are included in Section 2118 of the Organic Foods Production Act of 1990 (OFPA). Specifically,
280 the OFPA states the National List may allow the use of substances that would otherwise be prohibited
281 under organic regulations (i.e., synthetics) if the substance contains an active ingredient in the following
282 categories: “copper and sulfur compounds; toxins derived from bacteria; pheromones, soaps, horticultural
283 oils, fish emulsions, treated seed, vitamins and minerals; livestock parasiticides and medicines and
284 production aids including netting, tree wraps and seals, insect traps, sticky barriers, row covers and
285 equipment cleansers” (OFPA 2118(c)(B)(i)).

286 The NOP final rule currently allows the use of vitamins B₁, C and E as plant or soil amendments in organic
287 crop production under 7 CFR §205.601(j)(8). Synthetic forms of vitamins—including vitamins B₁, C and E—
288 are also permitted in other areas of organic production and handling. In addition to crop applications,
289 nutrient vitamins are also allowed as feed additives in organic livestock production under 7 CFR
290 §205.603(d)(3) in amounts needed for nutrition and health maintenance (7 CFR 205.237). Similarly,
291 synthetic sources of vitamins may also be incorporated into processed products labeled as “organic” or
292 “made with organic (specified ingredients or food group(s))” (7 CFR 205.605(b)).

293 **International**

294 Only Canadian organic regulations mention the use of synthetic vitamin compounds in organic crop
295 production. According to Section 4.2 of the Canadian General Standards Board’s Permitted Substances List,
296 synthetic forms of vitamins B₁, C and E are allowed soil amendments and crop nutrition agents in organic
297 agriculture (CAN, 2011). The following international standards and regulations were consulted but did not
298 contain guidance on the use of synthetic vitamins in organic crop production: Codex Alimentarius
299 Commission Guidelines for the Production, Processing, Labeling and Marketing of Organically Produced
300 Foods (CAC/GL 32-1999; Codex, 2013), European Commission Regulation Number 889/2008 (EC, 2008),
301 the IFOAM Norms for Organic Production and Processing (IFOAM, 2014), and the Japanese Agricultural
302 Standards for Organic Plants (Notification Number 1605; JMAFF, 2012).

303 **Evaluation Questions for Substances to be used in Organic Crop or Livestock Production**

304
305 **Evaluation Question #1: Indicate which category in OFPA that the substance falls under: (A) Does the**
306 **substance contain an active ingredient in any of the following categories: copper and sulfur**
307 **compounds, toxins derived from bacteria; pheromones, soaps, horticultural oils, fish emulsions, treated**
308 **seed, vitamins and minerals; livestock parasiticides and medicines and production aids including**
309 **netting, tree wraps and seals, insect traps, sticky barriers, row covers, and equipment cleansers? (B) Is**
310 **the substance a synthetic inert ingredient that is not classified by the EPA as inerts of toxicological**
311 **concern (i.e., EPA List 4 inerts) (7 U.S.C. § 6517(c)(1)(B)(ii))? Is the synthetic substance an inert**
312 **ingredient which is not on EPA List 4, but is exempt from a requirement of a tolerance, per 40 CFR part**
313 **180?**

314 (A) Vitamins B₁, C and E currently allowed for use as plant and soil amendments in organic crop
315 production fall under the category of vitamins and minerals; thus, these synthetic substances are eligible
316 for consideration under OFPA. Vitamin B₁ (thiamine) is a sulfur-containing compound.

317 (B) The previous paragraph provides sufficient information to determine eligibility of the substance under
318 OFPA; however, the inert status of vitamins B₁, C and E is briefly described. Vitamin E and L-ascorbic acid
319 (vitamin C) appear on US EPA List 4A, minimum risk inert ingredients. Thiamine mononitrate and vitamin
320 B complex are present on List 4B, minimal risk other ingredients. Vitamin E acetate (a synthetic form of
321 vitamin E) is included on List 3, inerts of unknown toxicity.

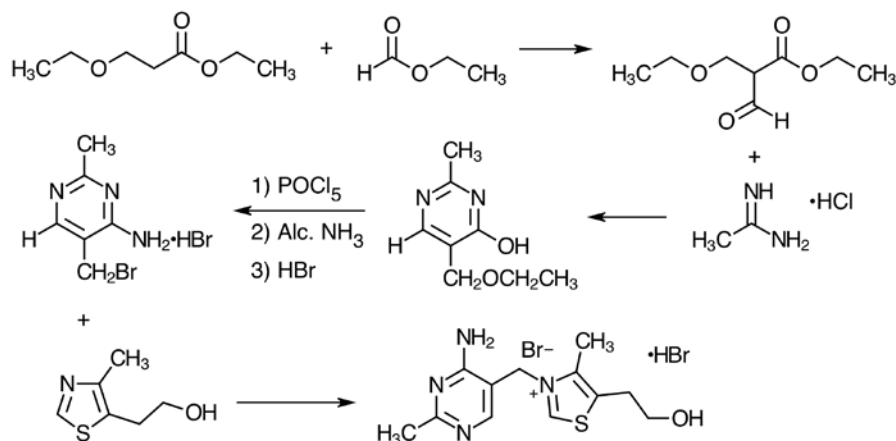
322 **Evaluation Question #2: Describe the most prevalent processes used to manufacture or formulate the**
323 **petitioned substance. Further, describe any chemical change that may occur during manufacture or**
324 **formulation of the petitioned substance when this substance is extracted from naturally occurring plant,**
325 **animal, or mineral sources (7 U.S.C. § 6502 (21)).**

326 Individual vitamin compounds are produced on an industrial scale by chemical synthesis or partial
 327 synthesis, fermentation and/or by extraction from natural material sources. Selection of the manufacturing
 328 processes typically depends on available technology, cost of raw materials/chemical feedstocks, market
 329 prices and size, cost of implementing fermentation versus chemical processes (synthesis or extraction) and,
 330 to a lesser extent, the overall environmental impact of the production method. Chemical synthesis is the
 331 primary production method for the three vitamins included in this technical review. Extraction of
 332 tocopherols (vitamin E) from plant oils is also commonly utilized, and laboratory-scale fermentation
 333 processes have been developed for both vitamin B₁ and C (Festel, 2005).

334 The following subsections summarize common manufacturing methods used in the commercial
 335 production of vitamins B₁, C and E. Processes reviewed in this section are provided as examples, and
 336 should not be considered the sole manufacturing procedures used for these vitamin compounds.

337 Vitamin B₁

338 Commercial production involves a six-step synthetic procedure (Williams & Cline, 1936). Beginning with
 339 ethyl 3-ethoxypropionate as the feedstock for vitamin B₁ production, the synthetic reactions include (1)
 340 formylation using ethyl formate, (2) reaction with acetamidine hydrochloride leading to aminopyrimidine
 341 ring formation, (3) replacement of aminopyrimidine hydroxyl group with a chlorine atom (chlorination)
 342 using phosphorus(V) oxychloride, (4) replacement of the labile chlorine atom with an amino group using
 343 alcoholic ammonia, (5) ammonium salt formation using hydrobromic acid, (6) introduction of the thiazole
 344 ring using 4-methyl 5-hydroxyethyl thiazole.



345
 346 **Scheme 1. The commercial production of thiamine salts (e.g., thiamine hydrobromide) involves a multi-**
 347 **step synthetic sequence (Adapted from Williams & Cline, 1936).**

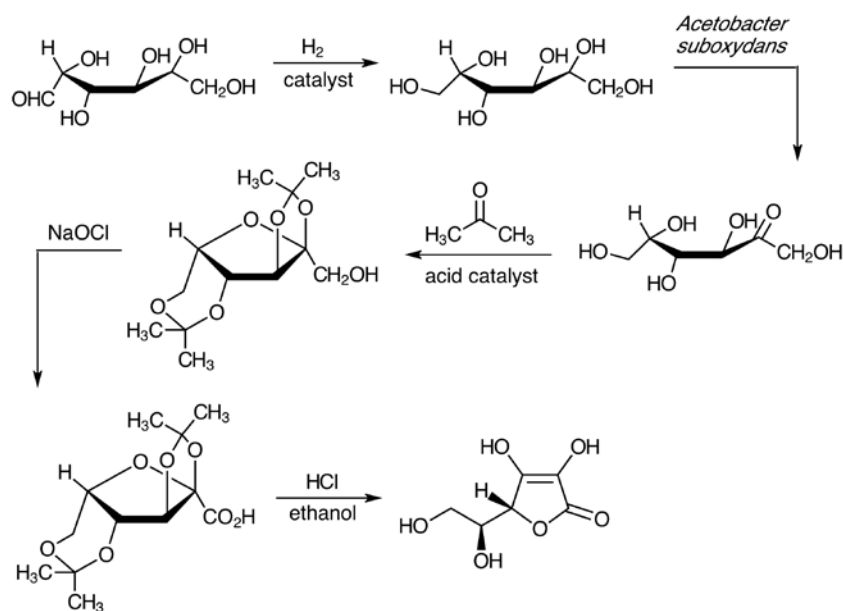
348 A search of the patent literature revealed two methods for vitamin B₁ (thiamine) production by
 349 fermentative methods. The first patent describes the development of mutants of the genus *Saccharomyces*
 350 *Meyen emend Reess* (yeast) for synthesizing vitamin B₁ from sugars and inorganic salts (Silhankova, 1980). A
 351 more recent invention provides a method for producing thiamine products using a microorganism of the
 352 genus *Bacillus* containing a mutation (i.e., gene deletions or other mutations) that causes it to overproduce
 353 and release thiamine products into the medium (Goese, 2012).

354 Vitamin C

355 Hoffmann-La Roche company synthesizes vitamin C from glucose through a five-step route. Glucose is
 356 first reduced to sorbitol using hydrogen and a transition metal catalyst. The microorganism *Acetobacter*
 357 *suboxydans* is then employed to oxidize sorbitol since no chemical oxidant is selective enough to oxidize
 358 only one of the six hydroxyl groups in sorbitol. Subsequent treatment with acetone and an acid catalyst
 359 converts four of the other hydroxyl groups into acetal linkages; the remaining hydroxyl group is
 360 chemically oxidized to the corresponding carboxylic acid through reaction with aqueous sodium
 361 hypochlorite (bleach). Hydrolysis with acid removes the two acetal groups and leads to an internal
 362 esterification yielding vitamin C (McMurry, 2011).

363 More recently developed synthetic strategies for producing vitamin C have also been described in the
 364 patent literature. One example involves the esterification of 2-keto-L-gulonic acid with a subsequent
 365 lactonization step and crystallization to form vitamin C (Fur, 1995). A related invention utilizes a similar
 366 synthetic process wherein L-ascorbic acid is produced in high yield through conversion of an aqueous
 367 solution of 2-keto-L-gulonic acid in the presence of an acid catalyst (Arumugam, 2003).

368 Recently a breakthrough fermentative method of vitamin C synthesis was disclosed, effectively
 369 transforming a 3-5 step chemical synthesis into a one-pot process (Festel, 2005). The patent literature also
 370 reveals a number of fermentative methods utilizing genetically modified microorganism for the
 371 overproduction of vitamin C (Beuzelin-Ollivier, 2012; Berry, 2001). The available information suggests that
 372 many vitamin C producing industries will ultimately shift toward fermentative methods using genetically
 373 modified microorganisms due to the increasing global demand for vitamin C and cost saving potential of
 374 these developing technologies (Festel, 2005).



375

376 **Scheme 2. The Hoffman-La Roche vitamin C commercial production involves five synthetic steps**
 377 **(Adapted from McMurry, 2011).**

378 *Vitamin E*

379 Synthetic vitamin E (α -tocopherol) is not identical to the form that occurs in nature; rather, it is a mixture of
 380 eight stereoisomers collectively known as all-rac-alpha-tocopherol, consisting of four 2R- and four 2S-
 381 isomers (Survase, 2006). Alternatively, a natural mixture of tocopherols can be extracted from vegetable oil
 382 sources (Vandamme, 1992). Extraction of tocopherols from vegetable oils typically involves a series of
 383 neutralization and separation stages following contact of the tocopherol containing substance with a
 384 caustic aqueous methanol solution and various aliphatic hydrocarbon solvents (Swanson, 1991).

385 Genetically modified organisms are potentially used in the production of vitamin E. Members of the
 386 organic community have voiced concerns over the use of vitamin E containing oils originating from
 387 genetically modified crop materials, particularly soybeans (Roseboro, 2008). In addition, a 2010 patent was
 388 filed for a vitamin E production process using biofermentation with algae or yeast that are genetically
 389 modified to enhance the production of farnesol and geranylgeraniol, potential starting materials in the
 390 syntheses of vitamins E and A (Maurina-Brunker, 2010).

391 **Evaluation Question #3: Discuss whether the petitioned substance is formulated or manufactured by a**
 392 **chemical process, or created by naturally occurring biological processes (7 U.S.C. § 6502 (21)).**

393 According to USDA organic regulations, the NOP defines synthetic as "a substance that is formulated or
 394 manufactured by a chemical process or by a process that chemically changes a substance extracted from
 395 naturally occurring plant, animal, or mineral sources" (7 CFR 205.2). Vitamins B₁ and C are likely derived

396 synthetically or through a combination of chemical synthesis and fermentation methods. Alternatively,
397 vitamin E (mixture of tocopherols) is typically extracted from natural materials (e.g., vegetable oils) using
398 aliphatic hydrocarbon solvents and acid-base extraction methods. Chemical synthesis and extraction
399 techniques are typically considered chemical processes due to the application of synthetic chemical
400 reagents in these methods. Vitamins produced through biological fermentation may be considered non-
401 synthetic or synthetic, depending on the nutrient feedstocks, fermentation organisms, and processing aids
402 used during production. Based on the NOP definitions and the predominant manufacturing processes, it is
403 reasonable to conclude that vitamins B₁, C and E are synthetic substances. The NOSB classified vitamins B₁,
404 C and E as synthetic following the 1995 TAP review; hence, these vitamins were included on the list of
405 synthetic substances allowed for use in organic crop production (7 CFR 205.601).

406 **Evaluation Question #4: Describe the persistence or concentration of the petitioned substance and/or its**
407 **by-products in the environment (7 U.S.C. § 6518 (m) (2)).**

408 In the course of production, use and disposal, vitamins B₁, C and E will be released to soil and water. This
409 section summarizes information concerning the environmental fate and persistence of vitamins B₁, C and E
410 in terrestrial and aquatic environments.

411 Water-soluble vitamins, such as vitamin B₁ and C, are expected to have slight to high mobility if released to
412 soil and therefore may spread to other soil areas and waterways (HSDB, 2010a). In contrast to vitamin C,
413 vitamin B₁ is more likely to adsorb to clay-based soils and other soil types with significant amounts of
414 organic matter (Schmidhalter, 1994). Water-soluble vitamins are unlikely to volatilize from moist and dry
415 soils due to their high polarity and low vapor pressures, respectively (HSDB, 2010a; HSDB, 2010b).
416 Vitamins B₁ and C are not expected to persist in the environment; for example, vitamin C exhibited a
417 biodegradability of 97% after 5 days and 100% after 15 days when applied to activated sludge (IPCS, 2014).
418 If released to water, vitamin C is not expected to adsorb to suspended solids and sediments (HSDB, 2010b),
419 while vitamin B₁ may exhibit low to moderate adsorption in aquatic environments (HSDB, 2010a). For
420 many water-soluble vitamins, the presence of functional groups that hydrolyze means hydrolysis is
421 expected to be an important degradation process. In contrast, volatilization of vitamins C and B₁ from
422 water surfaces is less likely. Water-soluble vitamins generally have low bioconcentration factors (BCFs),
423 suggesting minimal potential for bioaccumulation in aquatic organisms (HSDB, 2010a).

424 Fat-soluble vitamin compounds – such as the mixture of tocopherols comprising vitamin E – are less polar
425 than water-soluble vitamins and thus practically insoluble in water. Because of this property, vitamin E is
426 unlikely to be mobile in soils (HSDB, 2006). It is unlikely that fat-soluble vitamin E would volatilize from
427 dry soil based on its relatively low vapor pressure (1.4×10^{-8} mm Hg at 25 °C). If released to water, fat-
428 soluble vitamin E will adsorb preferentially to sediments and other suspended solids present in the water
429 column due to its solubility properties. The synthetic acetate esters of tocopherols used in many
430 commercial supplements readily hydrolyze to liberate the biologically active tocopherol species (van
431 Henegouwen, 1995); however, these free tocopherols that comprise vitamin E lack hydrolysable functional
432 groups. Thus, hydrolysis is not expected to be the dominant degradation process for vitamin E. The
433 bioconcentration factor (BCF) for the acetic ester of vitamin E is 3.2, indicating significant accumulation in
434 organisms is not expected (BASF, 2005). As components of highly biodegradable natural oils (e.g., soybean
435 oil), it is unlikely that tocopherols comprising vitamin E will persist in the environment due to use of the
436 substance to protect plant tissues and promote plant growth in crop production.

437 Erosion of soils containing fertilizers and other plant growth substances will increase the rate at which
438 phosphates, nitrates and nutrients such as vitamins enter streams, rivers, lakes and coastal regions (Muir,
439 2012). Ultimately, the persistence of the given vitamin compound may not be of paramount concern for
440 situations in which there is a continuous supply of nutrients resulting from intensive agricultural activities.
441 Laboratory-scale aquaculture studies have suggested that the accumulation of nutrients, including
442 vitamins, in bottom sediments may encourage the growth of algal blooms and red tide species (Wu, 1995).

443 **Evaluation Question #5: Describe the toxicity and mode of action of the substance and of its**
444 **breakdown products and any contaminants. Describe the persistence and areas of concentration in the**
445 **environment of the substance and its breakdown products (7 U.S.C. § 6518 (m) (2)).**

446 It is unlikely that the use of vitamins B₁, C and E as seed treatments, soil applications, and foliar sprays is
447 harmful to treated plants or non-target plants and animals. Vitamins are generally considered non-toxic
448 essential nutrients for terrestrial and aquatic organisms. Indeed, plants and microorganisms naturally
449 produce vitamin B₁, C and E for various biological functions. Based on the low hazard profile of these
450 substances, the potential for toxicity in any exposed organism is dependent upon the solubility properties
451 of the vitamin and the amount of exposure. In animals, the water-soluble vitamins (B₁ and C) are rapidly
452 depleted in the absence of regular dietary intake, and appreciable quantities of these vitamins do not
453 accumulate in the animal body. Fat-soluble vitamin E is readily absorbed from the gastrointestinal tract
454 and stored in the animal's fatty tissues when dietary intake exceeds metabolic demands for the vitamin.

455 Animals excrete vitamins B₁ and C in urine and generally do not exhibit symptoms of toxicity following
456 ingestion of these substances. Based on our review of the scientific literature and the 1998 Institute of
457 Medicine chapter for thiamine, there are no reports of adverse effects from consumption of excess vitamin
458 B₁ through ingestion of food and supplements. A tolerable upper intake level (UL) was not established for
459 thiamine due to the lack of hazard data necessary to conduct a quantitative risk assessment (Institute of
460 Medicine, 1998). There have been occasional reports of serious and even fatal anaphylactic reactions to the
461 administration of thiamine through routes other than the gastrointestinal tract. Allergic sensitivity and
462 itching were observed in 12 of 989 patients receiving intravenous injections of thiamine hydrochloride at
463 relatively high doses of 100 mg/day. These effects are irrelevant for setting a UL because the route of
464 exposure is not reflective of typical dietary intake for vitamin B₁ (Institute of Medicine, 1998).

465 Humans often take vitamin C supplements in large amounts because many people believe vitamin C is
466 non-toxic and beneficial to health. There is no evidence suggesting that vitamin C is carcinogenic or
467 teratogenic (i.e., causes developmental malformations) or that it causes adverse reproductive effects (i.e.,
468 fertility issues). Reviews of high vitamin C intakes have indicated low toxicity; however, adverse effects
469 have been reported after ingestion of doses exceeding three grams per day (Institute of Medicine, 2000).
470 The available data indicate that the human body does not continue to absorb dietary vitamin C at intakes
471 above 200 mg/day, suggesting that overload of vitamin C is unlikely in most humans. Very high intakes of
472 vitamin C may lead to diarrhea and other gastrointestinal disturbances, increased oxalate excretion and
473 kidney stone formation, increased uric acid excretion, pro-oxidant effects, systemic conditioning ("rebound
474 scurvy"), increased iron absorption leading to iron overload, reduced vitamin B₁₂ and copper status,
475 increased oxygen demand, and erosion of dental enamel (Institute of Medicine, 2000). Excessively high
476 vitamin C doses (e.g., up to 10 grams per day) may also interfere with the healthy antioxidant-prooxidant
477 balance in the body (Johnson, 2014a). Daily intake below the upper limit (UL = 2,000 mg/day) is not
478 associated with toxic effects in healthy adult humans (Institute of Medicine, 2000; Johnson, 2014a).

479 There is no evidence of adverse effects from consumption of vitamin E at levels naturally occurring in
480 foods. Instead, excessive intake of α -tocopherol in humans from supplementation, fortification of foods, or
481 pharmacological might reduce the coagulation properties of blood. This anticoagulant effect at high doses
482 could increase the risk of severe bleeding – reducing the blood's ability to form clots after a cut or injury –
483 or hemorrhagic stroke (Institute of Medicine, 2000). Animal studies have demonstrated that α -tocopherol is
484 not mutagenic, carcinogenic or responsible for fetal malformations (i.e., teratogenic). Occasionally, muscle
485 weakness, fatigue, nausea, and diarrhea occur at typical doses of α -tocopherol ranging from 400 to 800
486 mg/day (Johnson, 2014b). Bleeding is uncommon unless the dose is greater than 1,000 mg/day or the
487 patient takes oral coumarin or warfarin, which are commonly prescribed as anticoagulant agents. Based on
488 the hemorrhagic potential of vitamin E alone, an upper limit of 1,000 mg/day has been established for any
489 form of α -tocopherol in adults (Institute of Medicine, 2000; Johnson, 2014b).

490 The available hazard assessments for vitamins B₁, C and E pertain to effects associated with large doses of
491 vitamin supplements without consideration of metabolites. No sources were identified that discuss toxic
492 effects resulting from the breakdown products of either native vitamins B₁, C, and E or synthetic forms of
493 these vitamins commonly found in supplements and fortified foods. Further, no sources were identified
494 that discuss the possible persistence and areas of concentration of these vitamins compounds or their
495 breakdown products in the environment. Foliar residues of vitamin compounds could present moderate
496 toxicological risks for animals if target vegetation treated at excessively high application rates is ingested
497 very soon after treatment. However, the modest foliar application concentrations (e.g., 300 mg/L for

498 vitamin C) and rapid dissipation rates for these substances suggests that non-target animals are not at risk
499 of poisoning related to the current use pattern of vitamins in organic crop production.

500 **Evaluation Question #6: Describe any environmental contamination that could result from the**
501 **petitioned substance's manufacture, use, misuse, or disposal (7 U.S.C. § 6518 (m) (3)).**

502 The potential exists for environmental contamination resulting from the industrial production of several
503 vitamin compounds. Strong acids (e.g., hydrochloric acid and hydrobromic acid) used in the syntheses of
504 vitamins B₁ and C may alter the pH of aquatic systems if accidentally released to the environment. Strong
505 acids and bases are also utilized in the extraction of tocopherols from vegetable oils, and may lead to
506 environmental impairment if accidentally released or improperly handled. In addition, organic solvents
507 used in the commercial extraction of tocopherols from vegetable oil could result in environmental
508 contamination if released into the environment through waste streams. Many of the vitamins synthesized
509 for supplements and feed fortification are derived from petroleum products or genetically modified crop
510 materials. For example, acetone (CH₃COCH₃) used in the commercial synthesis of vitamin C is a high
511 volume production chemical derived from petroleum as well as genetically modified corn.

512 Waste streams resulting from the fermentative production of vitamins may also pose risks to the
513 environment. In general, the EPA assumes "no control features for the fermentor offgases, and no
514 inactivation of the fermentation broth for the liquid and solid waste releases," suggesting that
515 environmental exposure to these waste streams is highly likely (EPA, 1997). However, lacking are specific
516 examples of environmental damage resulting from exposure to recombinant DNA from genetically
517 modified microorganisms used in food and food additive production. Some potential risks to the
518 environment include the transfer of novel genes into crops, poisoned wildlife, the creation of new and
519 more potent viruses, as well as unanticipated health risks (UCS, 2002).

520 There is a slight risk of environmental contamination directly associated with the use of vitamins in organic
521 crop production. Chemical nutrients, such as vitamins, present in crop/soil amendments and livestock
522 feeds could be introduced to aquatic environments through accidental spills or leaching of nutrients from
523 treated soils. Some of these organic and inorganic nutrients have a propensity to accumulate in the bottom
524 sediments, which may lead to high sediment oxygen demand, anoxic sediments, production of toxic gases,
525 and a decrease in benthic diversity (Wu, 1995). However, it is unlikely that vitamins are the primary
526 drivers of environmental impairment due to their short half-lives in aquatic systems. Rather, laboratory
527 studies suggest that a continuous supply of vitamins may provide nutritional support to any algal blooms
528 and red tides that develop in eutrophic water bodies (Wu, 1995; NAS, 1969). Once algal proliferation
529 commences, available vitamins may therefore support the growing population. In particular, unicellular
530 photosynthetic algae require nutritional intake of vitamin B₁ (thiamine), among other B-vitamins (NAS,
531 1969). Therefore, a deficiency of these vitamins – as well as other macro- and micronutrients – can be a
532 limiting growth factor for environmentally beneficial and deleterious algae.

533 Overall, accidental releases of small amounts of vitamins into the environment are not assumed to pose
534 any significant risk. Material safety data sheets for many synthetic vitamins, including vitamin C, advise
535 that containers holding synthetic vitamins be "suitable" and closed containers for disposal. No further
536 disposal instructions are provided (Sigma Aldrich, 2015).

537 **Evaluation Question #7: Describe any known chemical interactions between the petitioned substance**
538 **and other substances used in organic crop or livestock production or handling. Describe any**
539 **environmental or human health effects from these chemical interactions (7 U.S.C. § 6518 (m) (1)).**

540 No direct chemical interactions between vitamins and other additives used in organic crop or livestock
541 production were identified. When ingested, vitamin C acts as a strong promoter of dietary iron absorption
542 while also counteracting the inhibitory effects of dietary phytate and tannins. Long-term vitamin C
543 supplementation may diminish the absorption of copper, thereby countering the beneficial effect on iron
544 absorption in humans. Further, there is evidence that vitamin C affects the bioavailability of selenium both
545 positively and negatively depending on the dietary conditions (Sandström, 2001). The synergistic
546 interaction of vitamin E and selenium as an oxidant defense system has been observed in a number of
547 animal species (Vannucchi, 1991). When vitamin E intercepts a radical to reduce a reactive oxygen species,
548 a tocopherol radical is formed; this radical can be reduced by ascorbic acid (vitamin C) to return vitamin E

549 to its biologically active state (Institute of Medicine, 2000). According to material safety data sheets, the
550 antioxidants vitamin C (L-ascorbic acid) and vitamin E (tocopherols) are incompatible with strong
551 oxidizing agents and alkali compounds (Sigma Aldrich, 2015; Sigma Aldrich, 2014).

552 **Evaluation Question #8: Describe any effects of the petitioned substance on biological or chemical**
553 **interactions in the agro-ecosystem, including physiological effects on soil organisms (including the salt**
554 **index and solubility of the soil), crops, and livestock (7 U.S.C. § 6518 (m) (5)).**

555 This technical evaluation report concerns the use of synthetic vitamins B₁, C and E as growth regulators
556 and plant protectants in organic crop production. It is highly likely that small amounts of these vitamins
557 would regularly interact with components of the terrestrial agro-ecosystem through runoff from treated
558 plants, leaching from soil, and accidental spills of vitamin solutions. Vitamins B₁, C and E – in addition to
559 numerous other vitamin species – are widely used in conventional and organic agriculture with no
560 reported toxicity observed in non-target wildlife or livestock.

561 No studies have been found indicating toxic effects of vitamins on soil-dwelling organisms. Some bacteria,
562 for example, do not require growth factors such as vitamins, while other bacterial strains (e.g., *Lactobacillus*)
563 require vitamins and other nutrients in order to grow (Todar, 2012). Therefore, despite the fact that some
564 water-soluble vitamins have the potential for high soil mobility, vitamins are unlikely to exhibit toxicity
565 toward the agro-ecosystem (HSDB, 2010a; HSDB, 2010b). Accidental release of chemical reagents during
566 industrial production, however, may lead to ecological impairment. Specifically, strong acids and bases are
567 used in the synthetic or extraction processes of vitamins B₁, C and E compounds (see response to
568 Evaluation Question #2 for details). Improper use or disposal of these chemicals during the production of
569 vitamins could affect both the pH and chemical composition of the soil, potentially resulting in
570 physiological effects on soil organisms. Reports of large-scale environmental releases or contamination
571 associated with the industrial production of vitamins were not identified.

572 **Evaluation Question #9: Discuss and summarize findings on whether the use of the petitioned**
573 **substance may be harmful to the environment (7 U.S.C. § 6517 (c) (1) (A) (i) and 7 U.S.C. § 6517 (c) (2) (A)**
574 **(i)).**

575 It is unlikely that the use of vitamins B₁, C and E as growth regulators and plant protectants in organic crop
576 production would be harmful to the environment. All photosynthetic organisms produce vitamins B₁, C
577 and E, and these nutrients are naturally part of the agro-ecosystem – from plant and animal tissues to the
578 soils supporting symbiotic, vitamin-producing microorganisms. Based on the presumably low application
579 rates and rapid dissipation, vitamin treatments as foliar applications, seed coatings and soil drenches
580 should not result in a substantial increase in the concentration of vitamins in the environment.

581 Strong acids (e.g., hydrochloric acid and hydrobromic acid) used in the syntheses of vitamins B₁ and C may
582 alter the pH of aquatic systems if accidentally released to the environment. Strong acids and bases are also
583 utilized in the extraction of tocopherols from vegetable oils, and may lead to environmental impairment if
584 accidentally released or improperly handled. Organic solvents used in the commercial extraction of
585 tocopherols from vegetable oil could also result in environmental contamination if released into the
586 environment through waste streams. However, no sources were identified that discussed environmental
587 contamination resulting from the manufacturing, transport or use of vitamins B₁, C and E.

588 **Evaluation Question #10: Describe and summarize any reported effects upon human health from use of**
589 **the petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (i), 7 U.S.C. § 6517 (c) (2) (A) (ii) and 7 U.S.C. § 6518**
590 **(m) (4)).**

591 In addition to being essential nutrients, vitamins are generally considered non-toxic and safe for human
592 consumption at levels typically ingested through the diet and dietary supplements taken according to label
593 directions. This response summarizes the available technical information regarding reported human health
594 effects associated with direct consumption of vitamins included in this review. Treatment of crops and soils
595 with vitamins B₁, C and E is unlikely to result in excessive vitamin intake for humans; hence, the
596 agricultural use pattern for these three vitamin species should not adversely affect impact human health.

597 *Vitamin B₁*

598 No adverse effects have been observed relating to the consumption of foods or dietary supplements
599 containing vitamin B₁ (thiamine). There have been occasional reports of anaphylaxis to parenteral thiamine
600 as well as pruritus due to allergic sensitivity to thiamine injection. A tolerable upper intake level (UL,
601 maximum level of daily nutrient uptake that is likely to pose no risk of adverse effects) was not determined
602 for thiamine due to the lack of data of adverse effects (Driskell, 2009; Institute of Medicine, 1998).

603 *Vitamin C*

604 Excess vitamin C intake is associated with osmotic diarrhea and gastrointestinal disturbances as the
605 primary adverse effects. Other possible effects include increased oxalate excretion and kidney stone
606 formation, increased uric excretion, pro-oxidant effects, rebound scurvy, increased iron absorption leading
607 to iron overload, reduced vitamin B₁₂ and copper levels, increased oxygen demand, and erosion of dental
608 enamel. UL for vitamin C is 2,000 mg/day (Driskell, 2009; Institute of Medicine, 2000).

609 *Vitamin E*

610 Adverse effects have not been observed from the consumption of vitamin E naturally occurring in foods.
611 High intakes of vitamin E from fortified foods, dietary supplements, or pharmacologic agents have
612 resulted in an increased tendency to hemorrhage as the primary adverse health effect. This anticoagulant
613 effect can be particularly severe for individuals deficient in vitamin K, including those taking coumarin
614 drugs. UL for vitamin E in the α-tocopherol form is 1,000 mg/day (Driskell, 2009; Institute of Medicine,
615 2000).

616 **Evaluation Question #11: Describe all natural (non-synthetic) substances or products which may be**
617 **used in place of a petitioned substance (7 U.S.C. § 6517 (c) (1) (A) (ii)). Provide a list of allowed**
618 **substances that may be used in place of the petitioned substance (7 U.S.C. § 6518 (m) (6)).**

619 Vitamins are available in a variety of natural plant and animal materials. In addition, various naturally
620 occurring, non-synthetic and synthetic substances applied as plant and soil amendments may perform
621 similar functions to those for vitamins B₁, C and E described in this report. The following sections provide
622 details regarding the natural availability of vitamins and alternative substances for root growth stimulation
623 (vitamin B₁) and antioxidant plant defense (vitamins C and E).

624 *Natural Sources of Vitamins B₁, C and E*

625 There are no direct substitutes for essential vitamins; however, plants readily synthesize vitamins B₁, C and
626 E within their roots and leaves for distribution throughout the entire plant. For example, plants generate
627 vitamin B₁ in the leaves and transport the compound to various plant tissues, including the root zone (Cox,
628 2010). Mycorrhizal fungi also secrete vitamin B₁ for plant uptake through the roots (Strzelczyk & Leniarska,
629 1985). Vitamins B₁, C and E are present in very small quantities within animal and plant foodstuffs. Natural
630 sources of these vitamins include the following:

- 631 • **Vitamin B₁:** Dried brewers yeast, wheat middlings, wheat mill run, rice bran, rice polishings, dried
632 torula yeast, groundnut (peanut) meal, wheat bran, barley, dried fish solubles, cottonseed meal,
633 soybean meal, linseed meal, dried distillers solubles, broad beans, lima beans, dried delactose
634 whey, glandular meals (liver/kidney), green leafy crops, outer coat or germ of cereals.
- 635 • **Vitamin C:** Citrus fruits, black currants, green leafy vegetables, green peppers, cauliflower,
636 watercress, green cabbage, strawberries, green cabbage, potatoes, fresh insects, and glandular
637 meals (liver/kidney).
- 638 • **Vitamin E:** Alfalfa meal, wheat germ meal, whole chicken eggs, rice polishings, rice bran, wheat
639 middlings, dried brewers grains, dried distillers solubles, barley grain, full fat soybean meal,
640 maize grain, wheat mill run, corn gluten meal, wheat bran, rye grain, sorghum, fish meal, oats,
641 sunflower seed meal, cotton seed meal, virtually all vegetable oils, and green leafy chops.

642 *Alternative Substances for Vitamins B₁, C and E in Crop Production*

643 Agricultural extension specialists generally recommend that conventional crop producers employ products
644 containing synthetic plant hormones or methods of stimulating the production of these hormones within
645 plants to enhance root growth in transplants and cuttings. Common synthetic auxins include indole butyric

646 acid (IBA) and naphthalene acetic acid (NAA). IBA is one of the most common auxin formulations, and has
647 been found to increase the number of roots and rooting percentage as well as regenerate roots in
648 transplanted trees (Chalker-Scott, undated). Neither synthetic IBA nor NAA is approved for use in organic
649 crop production (Schalau, 2010). Willow tea, made by soaking stems of the willow tree in warm water, is
650 often recommended for use as a natural source of the IBA hormone to encourage new cuttings to take root
651 (OSU, 2015); however, technical literature validating or refuting these claims is unavailable. A search of the
652 OMRI generic materials database indicates that nonsynthetic plant hormones such as gibberellic acid,
653 indole acetic acid (IAA) and cytokinins may be applied to organic crops as plant growth regulators (OMRI,
654 2013). Plant hormones regulate the cell division and cell elongation in general, but each group of plant
655 hormones performs specialized functions (Pederson, 2007). For example, auxins stimulate root growth,
656 gibberellins control flowering, and abscisic acid inhibits the effects of other hormones to reduce growth
657 during times of plant stress (Schalau, 2005; Whiting, 2010).

658 Strategies encouraging auxin formation within plants and applying natural substances/organisms
659 containing these hormones for root growth stimulation is most compatible with organic production.
660 Specifically, germinating seeds, fungi and algae extract may serve as natural auxin sources. Cuttings have
661 been enclosed with germinating seeds to stimulate root growth for several centuries because seeds
662 naturally produce auxins such as IAA when they germinate (Centeno & Gómez-del-Campo, 2008). A
663 commercial extract of macerated cereal seeds known as Terrabal Organico™ – which contains soluble
664 proteins, amino acids, vitamins, nitrogen, phosphorus and potassium – has been effectively used to
665 increase the percentage of rooted cuttings in organic crop production (Centeno & Gómez-del-Campo,
666 2008). Just as soil fungi naturally synthesize vitamin B₁, these same microorganisms generate auxins as well
667 as other essential plant nutrients, such as proteins, carbohydrates, lipids, minerals and other vitamins
668 (Centeno & Gómez-del-Campo, 2008). Encouraging the health of existing soil fungi and supplementing
669 soils with exogenous sources of beneficial fungi that release plant nutrients and growth factors to the soil
670 may naturally stimulate root growth in transplanted crops. Algae also contain IAA, proteins, lipids and
671 carbohydrates; for example, the Sm-6 Organico™ product is a mixture of *Ascophyllum nodosum*, *Fucus*
672 *serratus*, *Laminaria hyoborea*, and *Laminaria digitata* algae dry used to encourage root development in organic
673 cuttings (Centeno & Gómez-del-Campo, 2008). According to Centeno & Gómez-del-Campo (2008), the
674 organic products Terrabal (macerated cereal seeds) and Sm-6 (mixture of algae dry extract) produced the
675 highest rooting percentages in propagation studies of organic olive plants (*Olea europea* L. cv. Cornicabra).

676 A search of the Arbico Organics catalog revealed several naturally derived, OMRI-listed substances
677 marketed to stimulate root growth. Pumice is a lightweight volcanic rock that is widely used in growing
678 media such as soil and potting mixes to aid in aeration (i.e., allow airflow around plant roots), moisture
679 retention and to loosen heavy, clay-based soils (Arbico Organics, 2015a). The commercial product
680 Humboldt Roots is a concentrated liquid root stimulant that can be mixed with and applied along with
681 other organic fertilizers. It is derived from *Ascophyllum nodosum* seaweed and potassium humate, and
682 contains humic acid lignite and extract of *Quillaja saponaria*, commonly known as Soapbark (Arbico
683 Organics, 2015a). In addition, Superzyme® Biological Growth Factor Powder 1-0-4 is an organic fertilizer
684 containing plant food extracted from the fermentation of protein and *Bacillus spp.*, *Pseudomonas putida* and
685 *Trichoderma spp.* used to optimize absorption of water and nutrients (Arbico Organics, 2015a). Hydrolyzed
686 seaweed contains a wide range of micronutrients and plant-growth chemicals and hormones (e.g., auxins
687 and gibberellins) (Gush, 2011). In fact, OMRI-listed products such as Maxicrop Soluble Seaweed Powder
688 are marketed for “strengthening cuttings, bare root trees and shrubs with an application before and after
689 transplant” (Arbico Organics, 2015a). Mycorrhizal fungi that form symbiotic relationships with the rooting
690 systems of plants are also included in commercial formulations such as the Arbico Organics™ Root
691 Maximizer – Mycorrhizal Fungi product, which is designed for transplanted trees, vines and shrubs as well
692 as small transplants/seedlings and cuttings (Arbico Organics, 2015b).

693 No natural substances were identified as alternatives for the antioxidants vitamins C and E in organic crop
694 production. However, the utility of external sources of these substances is uncertain due to the paucity of
695 literature describing practical applications of these substances in agricultural settings.

696 **Evaluation Question #12: Describe any alternative practices that would make the use of the petitioned**
697 **substance unnecessary (7 U.S.C. § 6518 (m) (6)).**

698 From irrigation intensity to fertilization schedules, cultural practices can greatly influence crop growth and
699 development as well as the concentration of vitamins in plant tissues, including roots, leaves and fruit. This
700 response provides information regarding alternative methods for encouraging root growth in transplants
701 and cuttings and naturally increasing the vitamin content of plant tissues.

702 *Vitamin B₁*

703 Most agricultural experts agree that vitamin B₁ treatments do not reduce transplant shock or stimulate new
704 root growth in transplanted crops and plant cuttings grown outside of the laboratory (i.e., tissue culture
705 studies). Because beneficial soil fungi and bacteria associated with plant roots produce vitamin B₁ (Schalau,
706 2010), crop producers can foster the production of vitamin B₁ in the soil area by encouraging the growth
707 and productivity of beneficial soil microorganisms. Reducing fertilizer use is one potential strategy, since
708 fertilizers can have negative impacts on beneficial soil microorganisms such as mycorrhizal fungi, bacteria
709 and protozoa (Schalau, 2010). Application of fertilizers at the time of planting is therefore not
710 recommended by the University of Arizona agricultural extension. Proper irrigation has been shown to
711 help plant root systems become established successfully in more arid climates (Schalau, 2010). Operators
712 are encouraged to ensure that the applied irrigation not only saturates the root ball of a transplant, but also
713 some of the adjacent native soil. Proper saturation of the surrounding native soil encourages new roots to
714 expand and colonize a greater soil volume, which helps the plant better utilize available soil nutrients and
715 water resources (Schalau, 2010). Overall, the available literature does not support the premise that foliar
716 and soil applications of vitamin B₁ are responsible for root stimulation in transplanted crops (Schalau, 2010;
717 Cox, 2010; Chalker-Scott, Undated; Kontaxis & Cox, 1984).

718 Other factors that influence rooting in cuttings of herbaceous plants include health and condition of the
719 parent plant, timing, the rooting medium, water, light, and temperature. Using only vigorous, healthy
720 plants that are free of diseases and infestations as sources of cuttings can increase the likelihood of
721 successful rooting and plant propagation (Guse & Larsen, 2001). Early morning is typically the best time to
722 take cuttings from health plants because the plant is fully turgid (Evans & Blazich, 2015). The supporting
723 medium should provide the plant cutting with physical support, an adequate supply of oxygen and water
724 to the root zone, and proper drainage. In general, the best media for rooting cuttings combines two
725 materials: one that retains moisture and another with large pore spaces, which results in good aeration and
726 drainage (Guse & Larsen, 2001). Light is necessary for photosynthesis to provide the energy required to
727 form tissues that become roots and shoots; however, most thin-leaved plants react best if they are never
728 placed in direct sunlight (Guse & Larsen, 2001). Shading materials are commonly used to reduce natural
729 sunlight by 30 percent in areas with higher light intensity (Hamilton & Midcap, 2003). For optimum
730 rooting, it is best to maintain a temperature that encourages growth processes but does not cause excessive
731 moisture loss and wilting. The use of heating cables beneath the rooting medium allows the root zone to be
732 kept five to ten degrees warmer than air temperature, thus encouraging rooting without stressing higher
733 plant materials (Guse & Larsen, 2001).

734 *Vitamins C and E*

735 Light and average temperature have a strong influence on the chemical composition of horticultural crops
736 (Lee & Kader, 2000). While light is not essential for the synthesis of ascorbic acid in plants, the amount and
737 intensity of light during the growing season positively influences the amount of ascorbic acid formed. For
738 plant fruits, it has been observed that outside fruit exposed to maximum sunlight contain higher amounts
739 of vitamin C than inside and shaded fruit on the same plant (Lee & Kader, 2000). In general, lower light
740 intensity during growth translates to lower ascorbic acid content of plant tissues. Temperature also
741 influences the composition of plant tissues during growth and development. Using citrus as an example,
742 fruit grown under cool temperatures (i.e., 20–22 °C day, 11–13 °C night) generally contain more vitamin C
743 than fruit harvested from hotter areas with temperatures of 30–35 °C during the day and 20–25 °C at night
744 (Lee & Kader, 2000). Crops grown in milder climates with plenty of sunshine – such as the coastal areas of
745 central California – are less likely to require exogenous sources of vitamin C based on this analysis of
746 sunlight and temperature. In the absence of practical information from extension services, it remains
747 uncertain whether the foliar and soil applications of vitamin C are essential for crop production.

748 Certain cultural practices may enhance or diminish the levels of vitamin C in plant materials (Lee & Kader,
749 2000). In one study, application of gibberellins was beneficial to green tea quality, increasing the

750 concentration of vitamin C by 18%. The application of increasing amounts of nitrogen fertilizer typically
751 correlates with decreasing vitamin C levels in fruits and vegetables (Yasuor, 2013). In fact, the results of one
752 study indicated that increasing the amount of nitrogen fertilizer from 80 to 120 kg per hectare decreased
753 vitamin C by 7% in cauliflower (Lee & Kader, 2000). Alternatively, application of potassium fertilizer was
754 found to increase ascorbic acid content. Stefanelli et al. (2010) also found that the intensive fertilizer and
755 water use associated with industrial agriculture generally results in lower concentrations of essential
756 nutrients – such as flavonoids, carotenoids, glucosinolates and ascorbic acid (vitamin C) – in fruit and
757 vegetable crops. Indeed, less frequent irrigation can increase the concentrations of dietary fiber, vitamin C,
758 protein, calcium and other nutrients in several crops, including leeks and broccoli (Lee & Kader, 2000).
759 Based on this information, horticultural crops grown under lower nitrogen supply and less frequent
760 irrigation may be preferred due to the high concentrations of vitamin C and low concentrations of nitrate.

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